



UNIVERSITÉ DE LILLE
FACULTÉ DE MÉDECINE HENRI WAREMBOURG
Année : 2024

THÈSE POUR LE DIPLÔME D'ÉTAT
DE DOCTEUR EN MÉDECINE

**La relation entre l'exposition résidentielle à la pollution
atmosphérique et les micro-ARN circulants chez les adultes vivant
en zone urbaine du nord de la France**

Présentée et soutenue publiquement le 20 septembre à 14h
au Pôle Formation
par **Audrey HUBERT**

JURY

Président :

Monsieur le Professeur Philippe AMOUYEL

Assesseurs :

Monsieur le Docteur Jean-Marc LO GUIDICE

Madame la Docteur Victoria GAUTHIER

Directeur de thèse :

Monsieur le Docteur Luc DAUCHET

Avertissement

La Faculté n'entend donner aucune approbation aux opinions émises dans les thèses :
celles-ci sont propres à leurs auteurs.

Table des matières

Avant-propos	5
Résumé	6
Abstract	8
Liste des abréviations	10
Table des illustrations	11
Contexte	12
I. La pollution atmosphérique	12
II. L'épigénétique	12
III. Objectifs	14
Article.....	15
I. Introduction	15
II. Materials and methods	16
III. Results.....	23
IV. Discussion	28
V. Conclusion	35
Discussion	36
I. Conclusion sur les principaux résultats	36
II. Perspectives.....	37
Conclusion.....	39
Références	40
Annexes.....	51

Avant-propos

Cette thèse, réalisée sous la direction de Luc Dauchet, a donné lieu article publié en 2023 dans la revue Environment International : Audrey Hubert, Djamel Achour, Céline Grare, Gianni Zarcone, Manon Muntaner, Aghiles Hamroun, Victoria Gauthier, Philippe Amouyel, Régis Matran, Farid Zerimech, Jean-Marc Lo-Guidice, Luc Dauchet. *The relationship between residential exposure to atmospheric pollution and circulating miRNA in adults living in an urban area in northern France.* Environment International, Volume 174, 2023, 107913, ISSN 0160-4120, <https://doi.org/10.1016/j.envint.2023.107913> .

Ce travail a été soutenu par plusieurs organismes, notamment le Centre Hospitalier Universitaire de Lille, grâce à la subvention BonusH attribuée à l'équipe IMPECS. Le projet a également bénéficié du soutien financier du Conseil Régional Nord Pas-de-Calais et du Fonds Européen de Développement Régional (FEDER-Presage #36034) dans le cadre du programme CPER Institut de Recherche en Environnement Industriel (IRENI). En outre, ce travail a été soutenu par le gouvernement français à travers le Programme Investissement d'Avenir (I-SITE ULNE / ANR-16-IDEX-0004 ULNE), géré par l'Agence Nationale de la Recherche, dans le cadre du projet "Santé Environnement : du risque territorial au risque individuel", cofinancé par la Métropole Européenne de Lille. Il s'inscrit également dans les projets de recherche CLIMIBIO et ECRIN du CPER.

Résumé

Introduction : Les micro-ARN sont des facteurs de régulation épigénétique capables de réprimer l'expression des gènes cibles et pourraient jouer un rôle dans les effets de la pollution atmosphérique sur la santé. L'objectif de la présente étude en population générale était d'examiner l'association entre l'expression des micro-ARN et l'exposition résidentielle à long terme aux PM₁₀ et au NO₂.

Méthode : Nous avons inclus 998 participants adultes non-fumeurs issus de l'enquête transversale ELISABET (2010-2014) dans la région urbaine de Lille, en France. Les niveaux moyens annuels de pollution résidentielle ont été estimés à l'aide d'un système de modélisation de dispersion atmosphérique. Dix micro-ARN ont été sélectionnés sur la base des données disponibles dans la littérature, ainsi que deux micro-ARN de référence (miR-93-5p et miR-191-5p), et ont été quantifiés par RT-qPCR. Des modèles de régression linéaire multivariée ont été utilisés pour étudier l'association entre les micro-ARN et la pollution atmosphérique. Le seuil de signification statistique (après correction pour le taux de fausse découverte) a été fixé à $p < 0,1$.

Résultats : L'exposition annuelle moyenne entre 2011 et l'année d'inclusion était de $26,4 \pm 2,0 \mu\text{g}/\text{m}^3$ pour les PM₁₀ et de $24,7 \pm 5,1 \mu\text{g}/\text{m}^3$ pour le NO₂. Chaque augmentation de $2 \mu\text{g}/\text{m}^3$ d'exposition aux PM₁₀ était associée à une augmentation de 8,6 % (IC 95 % [3,1 ; 14,3]; $p_{\text{FDR}} = 0,019$) de l'expression de miR-451a. Une augmentation de $5 \mu\text{g}/\text{m}^3$ de l'exposition au NO₂ était associée à une augmentation de 5,3 % (IC 95 % [0,7 ; 10]; $p_{\text{FDR}} = 0,056$) de l'expression de miR-451a, une diminution de 3,6 % (IC 95 % [-6,1 ; -1,1]; $p_{\text{FDR}} = 0,052$) de l'expression de miR-223-

3p, une diminution de 3,8 % (IC 95 % [-6,8 ; -0,7]; $p_{FDR} = 0,079$) de l'expression de miR-28-3p, une diminution de 4,3 % (IC 95 % [-7,7 ; -0,8]; $p_{FDR} = 0,055$) de l'expression de miR-146a-5p et une diminution de 4,0 % (IC 95 % [-7,4 ; -0,4]; $p_{FDR} = 0,059$) de l'expression de miR-23a-5p. La différence entre les deux micro-ARN de référence miR-93-5p et miR-191-5p était également associée à l'exposition aux PM₁₀ et au NO₂.

Conclusion : Nos résultats suggèrent que les micro-ARN circulants pourraient être des biomarqueurs précieux pour évaluer les effets de la pollution atmosphérique.

Abstract

Introduction: MicroRNAs are epigenetic regulatory factors capable of silencing the expression of target genes and might mediate the effects of air pollution on health. The objective of the present population-based study was to investigate the association between microRNA expression and long-term, residential exposure to atmospheric PM₁₀ and NO₂.

Method: We included 998 non-smoking adult participants from the cross-sectional ELISABET survey (2010-2014) in the Lille urban area of France. The mean residential annual pollution levels were estimated with an atmospheric dispersion modelling system. Ten microRNAs were selected on the basis of the literature data, together with two housekeeping microRNAs (miR-93-5p and miR-191-5p) and were quantified with RT-qPCRs. Multivariate linear regression models were used to study the association between microRNAs and air pollution. The threshold for statistical significance (after correction for the FDR) was set to $p < 0.1$.

Results:

The mean annual exposure between 2011 and the year of inclusion was 26.4 ± 2.0 $\mu\text{g}/\text{m}^3$ for PM₁₀ and 24.7 ± 5.1 $\mu\text{g}/\text{m}^3$ for NO₂. Each 2 $\mu\text{g}/\text{m}^3$ increment in PM₁₀ exposure was associated with an 8.6% increment (95%CI [3.1; 14.3]; $p_{\text{FDR}} = 0.019$) in miR-451a expression. A 5 $\mu\text{g}/\text{m}^3$ increment in NO₂ exposure was associated with a 5.3% increment ([0.7; 10]; $p_{\text{FDR}} = 0.056$) in miR451a expression, a 3.6% decrement (95%CI [-6.1; -1.1]; $p_{\text{FDR}} = 0.052$) in miR-223-3p expression, a 3.8% decrement (95%CI [-6.8; -0.7]; $p_{\text{FDR}} = 0.079$) in miR-28-3p expression, a 4.3% decrement (95%CI [-7.7; -0.8];

$p_{\text{FDR}} = 0.055$) in miR-146a-5p expression, and a 4.0% decrement (95% CI[-7.4; -0.4]; $p_{\text{FDR}} = 0.059$) in miR-23a-5p expression. The difference between the two housekeeping microRNAs miR-93-5p and miR-191-5p was also associated with PM₁₀ and NO₂ exposure.

Conclusion: Our results suggest that circulating miRNAs are potentially valuable biomarkers of the effects of air pollution.

Liste des abréviations

miRNA (miR)	Micro-ARN
PM ₁₀	Particular matter with diameter < 10 µm (Particule en suspension de diamètre inférieur à 10 µm)
NO ₂	Dioxyde d'azote
FDR	False discovery rate (Taux de fausse découverte)
SD	Standard deviation (écart-type)
IQR	Interquartile range (écart interquartile)
BMI	Body mass index (Indice de masse corporelle)
HbA1c	Glycosylated hemoglobin (Hémoglobine glyquée)
HDL	High-density lipoprotein (Lipoprotéines à haute densité)
LDL	Low-density lipoprotein (Lipoprotéine à faible densité)
Ct	Cycle threshold (Seuil de cycle)
hs-CRP	High-sensitivity C-reactive protein (Protéine C-réactive à haute sensibilité)

Table des illustrations

Table 1: Characteristics of the study participants (n = 998).....	24
Table 2: Association between residential PM ₁₀ and NO ₂ and miRNA, using the level of the miRNA minus the average of the two housekeeping miRNAs	25
Table 3: Association between inflammatory biomarkers and miRNA, using the miRNA level minus the average of the two housekeeping miRNAs.....	27

Contexte

I. La pollution atmosphérique

La pollution atmosphérique a un impact considérable sur la santé humaine. En 2019, l'OMS estimait que la pollution atmosphérique était responsable de 4,2 millions de décès prématurés chaque année (1).

Les particules fines, les oxydes d'azote et d'autres polluants présents dans l'air sont associés à une augmentation de la mortalité et de la morbidité à travers le monde, allant des maladies pulmonaires chroniques et des maladies cardiovasculaires à des troubles métaboliques et divers cancers (2–4).

Parmi les nombreux polluants atmosphériques les particules fines constituent un problème majeur. Ces particules, comme les PM₁₀, d'un diamètre inférieur à 10 micron, peuvent pénétrer profondément dans les voies respiratoires et entrer dans la circulation sanguine, déclenchant des réponses inflammatoires qui perturbent le fonctionnement normal des organes (5,6).

Les recherches continuent d'explorer les voies biologiques spécifiques par lesquelles la pollution atmosphérique influence la santé, notamment à travers l'analyse de biomarqueurs d'inflammation et de modifications épigénétiques, afin de mieux comprendre et atténuer ces impacts sanitaires.

II. L'épigénétique

L'épigénétique est l'étude des mécanismes qui modifient de manière réversible, transmissible et adaptative l'expression des gènes sans en changer la séquence d'ADN. Contrairement à la génétique, où les mutations affectent directement le code ADN et sont généralement irréversibles, les changements

épigénétiques sont réversibles et peuvent être influencés par divers facteurs environnementaux (7). Les principaux mécanismes épigénétiques comprennent la méthylation de l'ADN, les modifications des histones et les ARN non codants, notamment les micro-ARN (miARN).

Les micro-ARN sont de petits ARN non codants, simple brin, qui jouent un rôle crucial dans la régulation de l'expression des gènes en inhibant leur traduction en protéines (8). En se liant aux ARN messagers (ARNm), les micro-ARN inhibent leur traduction en protéines, ce qui influence divers processus biologiques clés tels que la croissance cellulaire, la différenciation, l'apoptose et le métabolisme (8). Cette capacité à moduler l'expression de nombreux gènes rend les micro-ARN cruciaux dans la régulation de fonctions cellulaires normales et pathologiques (9).

Des dérégulations des micro-ARN sont associées à une gamme étendue de maladies, y compris les cancers, les maladies cardiovasculaires et les troubles auto-immuns (9–11). En outre, les micro-ARN jouent un rôle clé dans la régulation de la structure de la chromatine et de l'expression génique, influençant des processus tels que la tumorigenèse (12).

Leur potentiel en tant que biomarqueurs épigénétiques et cibles thérapeutiques représente un domaine de recherche prometteur (13–15). En affectant l'expression de gènes critiques, les micro-ARN offrent des perspectives significatives pour le diagnostic et le traitement de diverses maladies.

En résumé, l'épigénétique aide à comprendre comment des facteurs externes comme la pollution atmosphérique peuvent avoir des impacts à long terme sur la santé humaine en modifiant l'expression des gènes. Les micro-ARN, qui jouent un rôle crucial dans la régulation épigénétique, sont particulièrement importants dans ce contexte.

III. Objectifs

L'objectif principal de notre étude était d'étudier la relation entre l'exposition résidentielle à long terme, de deux polluant atmosphériques ; les PM₁₀ et le NO₂, et les niveaux circulants de dix micro-ARN chez des habitants de la région lilloise.

L'objectif secondaire était d'étudier la relation entre la présence de biomarqueur de l'inflammation et les micro-ARN circulants.

Article

I. Introduction

Air pollution has a major impact on health in general and on lung, cardiovascular and metabolic diseases in particular (12,16). The association between air pollution and health is probably mediated by a number of biological pathways, which can be investigated by monitoring biomarkers of epigenetic modification or inflammation (17).

Epigenetic mechanisms have attracted increasing research interest in recent years because they can explain the interactions between environmental exposures and health-related events. Epigenetics can control gene expression through various mechanisms, ranging from structural changes in chromatin to post-transcriptional regulation. MicroRNAs (miRNAs) constitute a large family of small, non-coding post-transcriptional regulators that silence the expression of target genes by pairing with the cognate mRNAs, which leads to mRNA degradation or the abrogation of translation. Given that miRNAs modulate the expression of more than 60% of the genome, they regulate a wide variety of biological processes and are believed to be involved in the pathogenesis of many diseases, including cancer (9).

The expression of miRNAs can be tissue- and organ-specific (18). However, highly stable protein-bound miRNAs have also been detected in the majority of biological fluids, and particularly in the bloodstream (19–22). These circulating miRNAs might be either actively secreted by cells as mediators for intercellular and interorgan communication or passively released from dead cells after apoptosis, necrosis, tumor growth or tissue damage (19,23).

Several studies have reported that environmental exposure can induce alterations in the miRNA profile (24–28). Given that the level of miRNA expression

varies from one tissue to another, the profile of circulating miRNAs might reflect the specific responses of tissues exposed to a harmful environment (29). Circulating miRNAs (whether actively secreted or passively released) are therefore promising new non-invasive biomarkers of tissue damage (29).

The association between air pollution and miRNA has been only investigated in studies of small numbers of participants. The number of adults studied ranged from 14 (30) to 153 (31), and one study included 273 children (32). Furthermore, there are few published data on the association between miRNAs and long-term exposure to air pollution. Hence, the objective of the present study was to evaluate the association between mean long-term (annual) residential exposure to PM₁₀ and NO₂ and levels of a number of miRNAs thought to be modified by air pollution and to mediate the impact of air pollution on health.

II. Materials and methods

1- Study population

We included male and female non-smoking adults (aged from 40 to 64) having resided in the Lille urban area of northern France for the previous 5 years. These adults were participating in the population-based, cross-sectional Enquête Littoral Souffle Air Biologie Environnement (ELISABET) epidemiological population survey, conducted between 2010 and 2013 in the Dunkirk and Lille areas. The study's methodology has been described in detail elsewhere (16,17,33). Briefly, the participants were selected from electoral rolls by random sampling, with stratification for sex, age and city area (Lille or Dunkirk), and were contacted in random order. Data were collected at home or (occasionally) during a consultation in a healthcare establishment. A blood sample was collected during the same visit. We excluded

pregnant women, incarcerated individuals, people under legal guardianship, people unable to give their informed consent, and people lacking French social security coverage. In compliance with the French legislation on biomedical research, the study protocol was approved by the local investigational review board (*CPP Nord Ouest IV, Lille, France; reference: 2010-A00065-34; ClinicalTrials.gov identifier: NCT02490553*). All the participants provided their written, informed consent to participation in the study.

With a view to increasing the study's statistical power for a small number of subjects (thus limiting the cost of analysis), we included participants from the Lille urban area only because the standard deviation of the residential air PM₁₀ level was higher in Lille than in Dunkirk (1.936 µg/m³ versus 0.999 µg/m³, respectively). To limited missing data and enable subsequent genetic studies, we included participants of Caucasian origin with data on genetic variables. Lastly, we excluded individuals without acceptable spirometry data and those lacking a fasting blood sample for the biomarker assay.

2- Air pollution measurements

The estimation of residential air pollution exposure has been described previously (16,34,35). In brief, the annual mean concentrations of PM₁₀ and NO₂ over the period from 2010 to 2013 were taken from estimates produced by ATMO–Nord Pas de Calais, using an atmospheric dispersion modelling system. The latter incorporated meteorological, topographic and land-use data, pollutant emissions from natural sources and those related to human activity, and ambient air pollution data from monitoring stations. Concentration maps for PM₁₀ and NO₂ in Lille are shown on the ATMO website (36). We applied a spatial resolution of 25 × 25 m. Each

participant's place of residence was located within a 25 m grid. We assessed the annual exposure levels at the place of residence as the mean value of the four closest points in the grid, weighted by the inverse square distance to each point (16,35). The residential exposure was defined as the mean annual exposure at the residential address from 2010 to the year of inclusion.

3- miRNAs of interest

In the present study, we assayed eight circulating miRNAs in plasma prepared from an EDTA blood sample from each study participant: miR21-5p, miR-28-3p, miR-30b-5p, miR-126-3p, miR-132-3p, miR-146a-5p, miR-222-3p, and miR-223-3p. These miRNAs were selected on the basis of the literature data because they are reportedly deregulated after exposure to various environmental air pollutants and/or in patients with lung disease possibly related to environmental exposure. Indeed, previous studies reported changes in the expression of miR-21-5p, miR-28-3p, miR-30b-5p, miR-126-3p, miR-132-3p, miR-146a-5p, miR-222-3p, and miR-223-3p following exposure to particulate matter of various aerodynamic diameter (coarse, fine or ultrafine particles), and/or to other gaseous pollutants related to traffic or industrial/biomass combustion(26,31,37–43). On the other hand, the link between the deregulation of miR-21-5p, miR-30b-5p, miR-126-3p, miR-146a-5p, miR-222-3p or miR-223-3p and the pathogenesis chronic obstructive pulmonary disease (COPD), asthma, idiopathic pulmonary fibrosis, cystic fibrosis or lung cancer has also been described (43–48). Four additional miRNAs were assayed: miR-23a-3p and miR-451a as controls for hemolysis and blood cell contamination (49,50), and miR-93-5p and miR-191-5p as endogenous reference genes for data normalization, as described previously (51).

The stability of these endogenous miRNAs was confirmed using NormFinder software (52).

4- miRNA assays

For miRNA extraction, 200 μ L of plasma samples were incubated for 5 min at room temperature with five volumes of Qiazol Lysis Reagent (Qiagen, Courtaboeuf, France) containing 1 μ g of the extraction enhancer MS2 bacteriophage RNA (Sigma-Aldrich, Saint Quentin Fallavier, France). After mixing with 200 μ L of chloroform and incubation for 2 to 3 min at room temperature, the samples were centrifuged at 12,000 g for 15 min at 4°C. The RNAs were then extracted from the aqueous phase using the miRNeasy Serum/Plasma kit and the QIAcube instrument, according to the supplier's instructions (Qiagen). The purity and quantity of RNAs were assessed by spectrophotometric analysis (the Spark multimode microplate reader equipped with the NanoQuant Plate™; Tecan, Mannedorf, Switzerland).

Circulating miRNAs were reverse-transcribed using 2 μ L of extracted RNAs and the TaqMan® Advanced miRNA cDNA Synthesis Kit, according to manufacturer's instructions (ThermoFisher Scientific, Courtaboeuf, France). Briefly, the synthesis of cDNAs started by 3' poly(A) tailing and 5' ligation of an adaptor sequence to extend the miRNAs at the 5' and 3' ends. The miRNAs were then reverse-transcribed and preamplified (using universal primers) to uniformly increase the amount of cDNA for each target while maintaining the relative differential levels. The universal primers recognize the universal sequences added to every miRNA and help to ensure the absence of amplification bias. All the target miRNAs were quantified using the TaqMan™ Fast Advanced Master Mix and the following TaqMan Advanced miRNA Assays: hsamiR-21-5p_477975; hsa-miR-23a-3p_478532; hsa-

miR-28-3p_477999; hsa-miR-30b-5p_478007; hsa-miR-93-5p_478210; hsa-miR-126-3p_477887; hsa-miR-132-3p_477900; hsa-miR-146a-5p_478399; hsa-miR-191-5p_477952; hsa-miR-222-3p_477982; hsa-miR-223-3p_477983; hsa-miR-451a_478107 (ThermoFisher Scientific). The samples were amplified using the QuantStudio™ 12 K Flex Real-Time PCR System (Applied Biosystems, ThermoFisher Scientific), and the results were calculated using the comparative cycle threshold (Ct) method and normalized against the signals detected for miR-93-5p and miR-191-5p.

5- miRNA analysis

Using the raw extraction values and the quartiles of the distribution of each miRNA, we determined an interval [min; max] outside of which values were considered to be outliers: $\text{min} = \text{quartile 1} - (1.5 * [\text{interquartile range (IQR)}])$ and $\text{max} = \text{quartile 3} + (1.5 * [\text{IQR}])$.

The lowest raw miRNA extraction values (expressed as $-\Delta\text{Ct}$) corresponded to a very high number of RT-PCR cycles and thus an almost undetectable amount of miRNA. We replaced the low outlier values by reassigning them with the corresponding min value.

The presence of hemolysis or red blood cell contamination in the sample was evaluated in two ways. Firstly, four medical biologists independently determined the presence or absence of hemolysis by eye. Secondly, we calculated the difference between the levels of the two hemolysis control miRNAs (miR-23a-3p and miR-45a) (50). If the difference was greater than five, the sample was potentially contaminated by red blood cells. If the difference was greater than seven, the samples were considered to be hemolyzed.

We calculated the extraction values (normalized using the ΔCt method) by subtracting the mean value of the two housekeeping miRNAs [miRNA – mean (miR-93-5p and miR-191-5p)] from the value for each miRNA. The relative expression of the miRNAs (i.e. the variation between the different samples) was expressed as $2^{-\Delta\text{Ct}}$.

6- Inflammatory biomarker assays.

The inflammatory biomarker assays have been described previously (17). Briefly, the serum concentration of high-sensitivity C-reactive protein (hs-CRP) was measured in a nephelometric assay (BN ProSpec System, Siemens) with a detection range of 0.17 to 10 mg/L. Values below the limit of detection (LOD) were computed as $0.12 \text{ (LOD}/\sqrt{2}) \text{ mg/L}$. For cytokine measurements, we used MesoScale Discovery[®] electrochemiluminescent immunoassays (Meso Scale Diagnostics LLC, Rockville, MA, USA): multiplex assays for interleukin (IL)-1 β , IL-6, IL-8, IL-10, IL-17A, and tumor necrosis factor alpha (TNF α), and an individual assay for IL-22. Values below the lower limit of quantification (the lowest concentration on the standard curve which gave a percentage coefficient of variation of <20%) were counted as real concentrations.

7- Statistical analysis

We used multivariate linear regression models to estimate the effect of PM₁₀ and NO₂ exposure on the expression levels of selected miRNAs (ΔCt). We adjusted our models for age, sex, body mass index, and year of inclusion. We adjusted for BMI because obesity is strongly related to many metabolism pathways including inflammation (53). We have adjusted for the year of inclusion because the

construction of the exposure variable was strongly dependent of the inclusion year which may be associated to temporal confusion bias.

Effect estimates and their 95% confidence intervals were expressed as a percentage of the change in miRNA expression level for an increment in a defined unit of pollutant or biomarker, using the equation $(2^{(-\text{estimate coefficient} \times \text{range of increment})} - 1) \times 100$. The increments of 2 $\mu\text{g}/\text{m}^3$ and 5 $\mu\text{g}/\text{m}^3$ were used because they were close to the IQR of exposure (17). We studied the associations between miRNAs (ΔCt) and biomarkers of inflammation: cytokines (IL1 β , IL-6, IL-8, IL-10, IL-17A, IL-22, TNF α , and hs-CRP).

Due to the large number of comparisons, we applied Benjamini-Hochberg false discovery rate (FDR) method to correct p-values for multiple comparisons. The threshold for statistical significance (after correction for the FDR) was set to $p < 0.1$.

We performed a stratified analysis on gender and test interaction for air pollution/gender interaction.

For the sensitivity analyses, we carried out the same analyses but excluded the hemolyzed samples determined by eye or by calculation. Lastly, we conducted sensitivity analyses using other miRNA normalization techniques: either by subtracting the mean of the three most stable miRNAs (miR-93-5p and miR-191-5p and miR-23a-3p) or the mean of all the other miRNAs from the level of each individual miRNA. The normalization method based on the mean of all the other miRNAs usually requires the measurement of dozens of miRNA and so could not be applied to our main analyses; however, we felt that its use could be justified in the sensitivity analysis (with 12 miRNAs measured (54)).

All statistical analyses were performed using R software (version 4.2.0).

III. Results

1- Characteristics of the study population

The ELISABET study included 1668 participants in the Lille urban area, of whom 1335 were nonsmokers (either never-smokers or people having stopped smoking at least 12 months previously). We selected the 1125 participants estimated to be Caucasian on the basis of the available genetic data. We then excluded 72 participants without acceptable spirometry data and 26 participants who had not fasted before the blood sample collection. Ultimately, biomarkers could be assayed for 1004 of the remaining participants. Measurements of the normalization housekeeping miRNAs (miR-93-5p or miR-191-5p) were missing for 6 participants, leaving 998 in the main analysis (454 (45.5%) men; median age: 54) ([Table 1](#)). In this sample, four values were missing for miR-28-3p, 6 for miR-132-3p and 5 miR-30b-5p. There were no missing data for the other miRNAs.

Between 2011 and the year of inclusion, the mean PM₁₀ exposure was 26.4 ± 2.0 $\mu\text{g}/\text{m}^3$ and the mean NO₂ exposure was 24.7 ± 5.1 $\mu\text{g}/\text{m}^3$. The correlation between PM₁₀ and NO₂ exposure levels was strong ($r_{\text{pearson}} = 0.79$; 95%CI [0.77; 0.81]; $p = < 2.2\text{e-}16$). The mean and Sd of fold log₂ fold change of the levels of miRNA are reported in [supplemental table 1](#).

Table 1: Characteristics of the study participants (n = 998)

Variable	value
Age (year)	53.8 (7.2)
Male gender, n (%)	454 (45.5)
Height (cm)	169.6 (8.9)
BMI (kg/m ²)	26.8 (5.0)
Mean annual levels of PM ₁₀ pollution between 2011 and the year inclusion (µg/m ³)	26.4 (2.0)
Mean annual levels of NO ₂ pollution between 2011 and the inclusion year (µg/m ³)	24.7 (5.1)
hs-CRP (mg/L)	0.9 (0.5 ; 1.9)
IL1b (pg/mL)	0.0 (0.0 ; 0.1)
IL6 (pg/mL)	0.4 (0.3 ; 0.7)
IL8 (pg/mL)	2.7 (2.1 ; 3.8)
IL10 (pg/mL)	0.2 (0.1 ; 0.3)
IL17a (pg/mL)	0.8 (0.5 ; 1.3)
IL22 (pg/mL)	0.2 (0.1 ; 0.4)
TNFα (pg/mL)	2.4 (1.9 ; 2.9)
DBP (mmHG)	126 (115 ; 139.4)
HbA1c (%)	5.5 (5.2 ; 5.9)
Glycemia (g/L)	0.9 (0.9 ; 1)
Total cholesterol (g/L)	2.2 (1.9 ; 2.5)
Triglycerides (g/L)	0.9 (0.7 ; 1.3)
HDL (g/L) median (IQR)	0.6 (0.5 ; 0.7)
LDL (g/L) median (IQR)	1.4 (1.2 ; 1.6)

Data are expressed as the mean (SD), the median [IQR] or n (%)

BMI: body mass index, PM₁₀: particulate matter < 10 µm, NO₂: nitrogen dioxide, HbA1c: glycosylated hemoglobin, HDL: high-density lipoprotein, LDL: low-density lipoprotein.

2- Main analysis: the association between air pollution exposure and miRNA expression

After correction for the FDR ($p_{FDR} < 0.1$), we found that one miRNA (miR-451a) was positively associated with both PM₁₀ and NO₂ levels and that four miRNAs levels (miR-223-3p, miR-28-3p, miR-146a-5p, and miR-23a-5p) were negatively associated with the NO₂ level ([Table 2](#)). A 2 µg/m³ increment in PM₁₀ exposure was associated with an 8.6% increment (95% confidence interval [3.1; 14.3]; $p_{FDR} = 0.019$) in miR451a expression ([Table 2](#)). A 5 µg/m³ increment in NO₂ exposure was associated with a 5.3% increment ([0.7; 10]; $p_{FDR} = 0.056$) in miR-451a and a 3.6% decrement ([-

6.1; - 1.1]; $p_{FDR} = 0.052$) in miR-223-3p, a 3.8% decrement ([-6.8; - 0.7]; $p_{FDR} = 0.079$) in miR-28-3p, a 4.3% decrement ([-7.7; - 0.8]; $p_{FDR} = 0.055$) in miR-146a-5p, and a 4.0% decrement ([-7.4; - 0.4]; $p_{FDR} = 0.059$) in miR-23a-5p (Table 2). We also observed that the difference between the two housekeeping miRNAs (miR-93-5p normalized against miR-191-5p or vice versa) were significantly associated with PM₁₀ and NO₂.

Table 2: Association between residential PM₁₀ and NO₂ and miRNA, using the level of the miRNA minus the average of the two housekeeping miRNAs

micro-RNA	PM ₁₀ (per 2 µg/m ³ increment)			NO ₂ (per 5 µg/m ³ increment)		
	% difference [95CI%]	p	p FDR*	% difference [95CI%]	p	p FDR*
miR.451a	8.6 [3.1 ; 14.3]	0.002	0.019	-3.6 [-6.1 ; -1.1]	0.005	0.052
miR.28.3p	-4.3 [-7.8 ; -0.6]	0.022	0.112	-3.8 [-6.8 ; -0.7]	0.016	0.079
miR.223.3p	-3.1 [-6 ; -0.1]	0.042	0.140	-4.3 [-7.7 ; -0.8]	0.016	0.055
miR.132.3p	3.1 [-0.8 ; 7.2]	0.117	0.292	5.3 [0.7 ; 10]	0.022	0.056
miR.146a.5p	-3.0 [-7 ; 1.2]	0.164	0.328	-4.0 [-7.4 ; -0.4]	0.029	0.059
miR.23a.5p	-3.0 [-7.1 ; 1.3]	0.166	0.277	-2.2 [-4.5 ; 0.2]	0.073	0.122
miR.126.3p	-2.0 [-4.7 ; 0.9]	0.174	0.248	-2.3 [-5.6 ; 1.1]	0.175	0.249
miR.222.3p	-1.6 [-4.9 ; 1.9]	0.362	0.452	-1.9 [-4.7 ; 1]	0.205	0.256
miR.30b.5p	-1.7 [-5.6 ; 2.3]	0.395	0.438	1.5 [-1.7 ; 4.9]	0.359	0.399
miR.21.5p	0.5 [-2.7 ; 3.7]	0.781	0.781	-0.8 [-3.4 ; 2]	0.577	0.577
miR.93.5p	3.8 [1.2 ; 6.4]	0.004		3.5 [1.3 ; 5.7]	0.002	
miR.191.5p	-3.6 [-6 ; -1.2]	0.004		-3.4 [-5.4 ; -1.3]	0.002	

* p value corrected for the false discovery rate

Adjusted for age, sex, body mass index, and year of inclusion

PM₁₀: particulate matter < 10 µm, NO₂: nitrogen dioxide

3- Sensitivity analyses

Stratified analysis on gender are presented in [supplemental table 2](#) and [3](#) none of the interaction test between air pollution and gender were Significant after FDR correction. miR-23a-5p, miR-223-3p and miR451a were significantly associated with NO₂ and PM₁₀ were significantly associated with miR-451a and miR-132-3p after

FDR correction in women. no association with NO₂ or PM₁₀ were significant in men.

A total of 130 (12.0%) samples with hemolysis were excluded: 26 by eye and 104 using the calculation method ([Supplemental Table 4](#)). Exclusion of the samples with hemolysis did not greatly modify the results, except that the association between PM₁₀ and miR-28-3p became statistically significant. We also observed fewer significant associations with NO₂ exposure after correction for the FDR.

When subtracting the mean level of the three most stable miRNAs (miR-93-5p, miR-191-5p and miR-23a-3p) from each miRNA, we found that two of the other miRNAs (miR-451a and miR-132-3p) were positively associated with PM₁₀ and one (miR-28-3p) was negatively associated with PM₁₀ ([Supplemental table 5](#)) and NO₂. However, only the association with miR-451a remained significant after correction for the FDR ([Supplemental Table 6](#)).

When subtracting the mean of all the other miRNAs from each miRNA, we found that three miRNAs were positively associated with PM₁₀ (miR-451a, miR-93-5p and miR-132-3p), three (miR-191-5p, miR28-3p,miR-223-3p) were negatively associated with PM₁₀ ([Supplemental Table 3](#)), three (miR-451a, miR-93-5p and miR-132-3p) were positively associated with NO₂, and five (miR-146a-5p, miR-23a-5p, miR-223-3p, miR-28-3p and miR-191-5p) were negatively associated with NO₂.

4- Additional analyses

Association between inflammatory biomarkers and miRNA. By considering the difference with the mean of the two-housekeeping miRNA, we found positive associations between inflammatory biomarkers and miRNAs. MiR-223-3p and miR-21-5p were associated with hs-CRP, IL-6 and IL-10; miR-30b-5p was associated with hs-CRP; miR126-3p was associated with IL-6 and IL-10; and miR-23a-5p and

miR132-3p were associated with IL-10. No significant associations were observed for IL-1 β , IL-8, IL-17a, TNF α , and IL-22.

Table 3: Association between inflammatory biomarkers and miRNA, using the miRNA level minus the average of the two housekeeping miRNAs.

	microRNA	% difference [95CI%]	p	p FDR*
hs-CRP	miR.223.3p	9.303 [3.1 ; 15.9]	0.003	0.028
	miR.21.5p	9.282 [2.8 ; 16.2]	0.004	0.022
	miR.30b.5p	10.545 [2.3 ; 19.5]	0.012	0.040
	miR.146a.5p	8.326 [-0.3 ; 17.7]	0.058	0.145
	miR.126.3p	5.334 [-0.3 ; 11.3]	0.063	0.126
	miR.28.3p	6.915 [-0.5 ; 14.9]	0.070	0.117
	miR.23a.5p	7.063 [-1.4 ; 16.3]	0.106	0.151
	miR.222.3p	4.344 [-2.4 ; 11.6]	0.213	0.267
	miR.451a	-5.755 [-14.8 ; 4.2]	0.250	0.277
	miR.132.3p	3.864 [-3.7 ; 12]	0.324	0.324
	miR.93.5p	-6,4 [-10,8 ; -1,7]	0.008	
	miR.191.5p	6,8 [1,8 ; 12,1]	0.008	
	IL6	miR.223.3p	7.067 [2.1 ; 12.3]	0.005
miR.21.5p		6.572 [1.4 ; 12]	0.013	0.064
miR.126.3p		5.690 [1.1 ; 10.5]	0.015	0.049
miR.30b.5p		4.958 [-1.5 ; 11.8]	0.133	0.332
miR.146a.5p		4.668 [-2.1 ; 11.9]	0.181	0.361
miR.23a.5p		3.916 [-2.8 ; 11.1]	0.263	0.438
miR.222.3p		1.607 [-3.8 ; 7.3]	0.566	0.809
miR.132.3p		0.329 [-5.5 ; 6.6]	0.915	1.144
miR.28.3p		0.216 [-5.5 ; 6.2]	0.942	1.047
miR.451a		0.091 [-7.8 ; 8.6]	0.983	0.983
miR.93.5p		-1,9 [-5,7 ; 2]	0.331	
miR.191.5p		2 [-1,9 ; 6]	0.331	
IL10		miR.23a.5p	26.498 [10.5 ; 44.8]	0.001
	miR.21.5p	14.571 [3.6 ; 26.7]	0.008	0.042
	miR.223.3p	13.134 [2.7 ; 24.6]	0.012	0.041
	miR.126.3p	10.429 [0.9 ; 20.8]	0.031	0.077
	miR.132.3p	13.045 [0.1 ; 27.7]	0.049	0.098
	miR.146a.5p	13.831 [-0.5 ; 30.3]	0.060	0.100
	miR.30b.5p	11.754 [-1.6 ; 26.9]	0.087	0.125
	miR.28.3p	10.364 [-1.9 ; 24.1]	0.101	0.126
	miR.451a	-8.640 [-22.6 ; 7.8]	0.284	0.316
	miR.222.3p	5.333 [-5.6 ; 17.6]	0.355	0.355
	miR.93.5p	-7,5 [-14,5 ; 0,1]	0.053	
	miR.191.5p	8,1 [-0,1 ; 17]	0.053	

* p value corrected for the false discovery rate.

hs-CRP: high-sensitivity C-reactive protein.

IV. Discussion

The results of our population-based study showed that long-term exposure to residential air pollution was associated with plasma miRNA levels and thus suggested that air pollution impacts gene regulation processes. Plasma levels of miR-223-3p, miR-28-3p, miR-146a-5p and miR-23a-5p were negatively associated with NO₂ exposure, and whereas the level of miR-451a was positively associated with NO₂ exposure. No significant interaction between gender and air pollution were observed. We didn't highlight any difference or air pollution effect between gender. When studying PM₁₀, only one association (with miR-451a) achieved statistical significance after correction for the FDR. Nevertheless, PM₁₀ and NO₂ were strongly correlated, and so it is hard to identify specific effects of one or the other. The standard deviations of NO₂ were higher than those of PM₁₀, which might have resulted in greater statistical power in the analysis of the association with NO₂.

Previous studies of the association between air pollution and these miRNAs had smaller samples and gave disparate results. These studies are described in [supplemental table 7](#). Most of previous studies focus on short term exposure: five studies had cross sectional design (25,37,38,54,55) including one with experimental exposure to diesel exhaust (54), six were small cohort with repeated measurement of exposure and miRNA (26,39,56–59) two were cross-sectional studies (32,60). Long term exposure was less studied, Only Rodosthenous et al (26) studied short- and long-term exposure (6 month and 1 year) with repeated measurement. The association with benzo[a] pyrene-r-7,t-8,t-9,c-10-tetrahydrotetrol and serum albumin(BPDE-Alb), a biomarker of PM exposure was studied in children (32) and a cohort of pregnant women studied exposure in pregnancy associated to miRNA in placenta at birth.

The negative association we observed between NO₂ and miR-23a-5p was consistent with negative association observed at short term with traffic air pollution (TRAP) in Chinese students (55) ([supplemental table 8](#)). But conversely to our findings a positive association for long term exposure were observed in the study of Rodosthenous et al (26). The negative association we observed between NO₂ and miR-28-3p has not been found in previous study. One positive association were observed at short term in a cross-sectional study (38). The negative association between NO₂ and miR-146a-5p were consistent with previous association observed with short term exposure (31,37,59) but conversely positive association at short term (26,38) and positive association with BPDE-Alb (32) were also observed. The negative association with miR-223-3p were inconsistent with positive association at short (26,54) and long term (26) observed previously. The negative association at long term we observed with miR-451a were inconsistent with the positive short-term association observed by Rodosthenous et al (26). Although miR-191-5p and miR-93-5p were used as housekeeping miRNAs the results suggest that at least one of the may be associated to air pollution, indeed Rodosthenous et al have previously found significant association between air pollution these miRNAs. Finally, Other associations have been observed for the miRNAs not associated to air pollution in our study and are listed in supplemental table 8 with biological effect or evaluated miRNAs. The inconsistency of result may be explained by difference population in population, exposure and methodology. More population based study with large sample are needed to confirm these results.

1- Physiological pathways

Inflammasomes are cytosolic protein complexes responsible for the activation

of inflammatory pathways in response to a wide range of pathogens and cell damage signals (55). Transcription of the most-studied inflammasome (NLR family pyrin domain containing 3 (NLRP3)) can be induced by Toll-like receptor (TLR) agonists and some inflammatory cytokines in an NF- κ B-dependent manner (56). Through activation of caspase 1, activation of the NLRP3 inflammasome allows the conversion of the proinflammatory cytokines IL-1 β and IL-18 into their mature forms, together with the induction of gasdermin-D-mediated pyroptotic cell death.

In order to avoid inappropriate activation, basal levels of inflammasome components are rather low in most cells. It is now well established that inflammasome functionalization can be modulated by a subset of miRNAs called inflammamiRs (57). Three of the miRNAs negatively correlated with pollution exposure in the present study are classified as inflammamiRs; miR-223-3p, miR-146a-5p, miR23a-3p.

MiR-223-3p was the first human miRNA found to directly target NLRP3 (58,59) and thus exhibits significant anti-inflammatory and antipyroptotic effects. In a number of studies, miR-223-3p expression is associated with exposure to multiple airborne contaminants. Nevertheless, the results of these studies are not always consistent and appear to vary as a function of the nature of the pollutant or the biological matrix analyzed. In a pilot study with 22 participants, Rodosthenous et al. (26) found a significant association between long-term ambient PM_{2.5} exposure and circulating levels of extracellular vesicle-encapsulated miR-223-3p. In accordance with these results, several studies have described increases in expression levels of this miRNA in response to other environmental challenges (42,61). In contrast, a significant downregulation of miR-223 was observed in human and rat lungs after tobacco smoke exposure (62,63). Lastly, in line with our present results, the plasma miR-223-

3p level was low in female CD-1 mice exposed to smoke condensate samples collected from different biomass burn scenarios (64).

As with miR-223, miR-146a has an anti-inflammatory role because it can downregulate the NF- κ B-mediated priming of the NLRP3 inflammasome. Indeed, miR-146a targets TLR4, myeloid differentiation primary response gene 88 (MyD88), interleukin-1 receptor-associated kinase 1 (IRAK1), and TNF-receptor-associated factor (TRAF6) – all key proteins in the canonical NF- κ B pathway (57,65). Moreover, miR-146a downregulates RelB, a pivotal factor in the non-canonical NF- κ B pathway (66,67). The anti-inflammatory role of miR-146a was also evidenced by the fact that miR-146a knock-out mice develop NF- κ B-driven low-grade inflammation (68,69). In agreement with our findings, several researchers have observed a negative correlation between PM exposure and miR-146a-5p levels (31,37,70,71).

In contrast, a 24-participant cross-over study of personal air pollution exposure assessment gave discordant results: the circulating level of miR-146a-5p was positively correlated with exposure to airborne ultrafine particles (38).

Lastly, miR-23a-3p can also help to downregulate the inflammasome by targeting the NLRP3-activating factors NIMA-related kinase 7 (NEK7) (72) or the CXCR4 chemokine receptor involved in stabilization of thioredoxin interacting protein (TXNIP) (73). Initially, miR-23a-3p was selected (along with miR-451a) to evaluate the level of hemolysis in biological samples. Very few studies have linked the expression of this miRNA with exposure to air pollution. In their pilot study of 22 participants, Rodosthenous et al. (26) observed a significant association between long-term ambient PM_{2.5} exposures and circulating miR-23a-3p levels. However, as with the two previously mentioned anti-inflammatory miRNAs (i.e. miR-223-3p and miR-146a-5p), we observed a negative correlation between the plasma miR-23a-3p

level and pollution exposure. Taken as a whole, these data suggest that organisms adapt to and control the inflammatory response induced by exposure to air pollutants. The association between short-term exposure to atmospheric air pollution and low-grade systemic inflammation previously observed in our study population might reflect this phenomenon (17).

Although not described as inflammamiRs, the two other miRNAs correlated with pollution exposure in our study (i.e. miR-28-3p and miR451a) regulate inflammatory signaling pathways. Yang et al. showed that miR-28 directly targets NRF2 (73), a major regulator of antioxidant and anti-inflammatory responses (74,75). Hence, miR-28-3p could be considered as a proinflammatory miRNA. In the present study, we observed a negative correlation between miR-28-3p levels and exposure to PM₁₀ and NO₂. Krauskopf et al.'s recent preliminary study of early biomarkers in only 30 participants showed that circulating miR-28-3p was positively correlated with a mixture of compounds found in traffic-related air pollution, including black carbon, CO₂, nitrogen oxides, and ultrafine particles (38). However, Krauskopf et al. did not find an association with PM_{2.5}.

Based on the literature data, miR-451a is a potential anti-inflammatory miRNA. Indeed, several studies have described how miR-451a inhibits the NF-κB pathway by reducing the expression of CAB39 (76), IKKB (77) and HMGB1 (78,79). Our present study evidenced a positive correlation between the plasma level of miR-451a and long-term pollution exposure. Motta et al. (79) did not find a significant association with PM₁₀ levels after 24 or 48 h of exposure. However, Motta et al.'s study included 90 overweight or obese adults, and obesity has already been shown to induce variability in the effects of PM (80,81). Similarly, Rodosthenous et al. found no association between short- and long-term PM_{2.5} exposure and the miR-451a level.

(26). In contrast, Deng et al. observed low miR-451a plasma levels in coke oven workers exposed to both metals and polycyclic aromatic hydrocarbons (82).

The tight control over inflammasome assembly and signaling is crucial for enabling the immune system to mount sufficiently strong antimicrobial and inflammatory responses to environmental challenges while avoiding the excessive tissue damage that could result from an uncontrolled reaction. Thus, deregulated inflammasome activation has been linked to the development of various diseases, from inflammatory diseases to cancers. In this context, any variation in the expression of inflammation-related miRNAs might reflect or cause pathological processes. It is therefore not surprising that the deregulation of the five miRNAs correlated with pollution exposure in our study has been linked to the pathogenesis of many diseases. In particular, the inflammasome modulating miRNAs miR-23a-3p, miR-146a-5p and miR-223-3p are reportedly dysregulated in a wide range of immune inflammatory diseases (such as multiple sclerosis, rheumatoid arthritis, systemic lupus erythematosus, inflammatory bowel disease, type 1 diabetes, and asthma (83,84), various non-immune inflammatory diseases (such as COPD and atopic, metabolic, neuroinflammatory and cardiovascular diseases (7,84–90), and several types of cancer (91–93). The expression of the remaining miRNAs (miR-28a-3p and miR-451a) is reportedly altered in tumoral diseases; these miRNAs have been suggested as promising cancer biomarkers with therapeutic potential (14,15,94). Lastly, abnormal levels of miR-451a have been observed in patients with cardiac fibrosis (94), cardiovascular diseases (95,96) and rheumatoid arthritis (97), and serum concentrations of miR-28a-3p are potential biomarkers in severe asthma (98) and COPD (99).

2- Strengths and weaknesses

The present study had several strengths. Firstly, to the best of our knowledge, this is the first population-based present study to have measured the association between air pollution and miRNAs. Secondly, our study included a large sample (n = 998) of non-smokers, which is more than in previous studies. Our study has greater sample size than previous ones it had enough power to detect association with similar effect size than those previously observed. For example, Fossati et al (31) observed a – 19.4% decrease of miR-126 for 3.02 $\mu\text{g}/\text{m}^3$ increase of 28 days exposure to PM_{2.5}. In our study the standard deviation of PM₁₀ exposure were 2 $\mu\text{g}/\text{m}^3$. to take in account that standard deviation(SD) of PM₁₀ which include PM_{2.5} are by definition higher than SD of PM_{2.5} we multiply the SD of PM₁₀ by 0.7 (16) and considering a SD of exposure at 1.4 $\mu\text{g}/\text{m}^3$ for PM exposure. The SD of difference of Ct between miRNA and average of the two housekeeping miRNAs were 0.604. according to these data with an alpha risk at 0.005 (to take in account FDR correction). the power our study was estimate at 98%. Thirdly, most previous studies looked at short-term exposure; we provided new data on long-term exposure with a fine-scale model (25x25m).

Our study also had some limitations. Firstly, we chose housekeeping miRNAs because of their known stability, which was confirmed here. Nevertheless, the observed association between housekeeping miRNAs and exposure might have influence the interpretation of our data. The association between housekeeping miRNAs and pollution can be explore indirectly only if several housekeeping miRNAs have been studied. This validity check is not generally described in the literature on house-keeping miRNA (28,51,100). Here, we identified a significant association

between air pollution and the difference between the two main housekeeping miRNAs; this prevented us from drawing any definite conclusions about the effect size of the observed associations. Nevertheless, our results are consistent because the associations observed in the main analysis were also seen in the sensitivity analyses.

V. Conclusion

We observed associations between residential long-term air pollution exposure and a number of miRNAs in large population-based sample of urban inhabitants. The miRNAs significantly associated with air pollution are mostly involved in inflammation suggesting that inflammation is a major pathway. Our results suggest that circulating miRNAs are potentially valuable biomarkers of the environmental risk.

Discussion

I. Conclusion sur les principaux résultats

Cette étude apporte des preuves supplémentaires quant à l'impact de la pollution de l'air à long terme sur la régulation génétique via les micro-ARN circulants. Les résultats montrent des associations significatives entre l'exposition résidentielle prolongée à des polluants atmosphériques, comme le NO₂ et les PM₁₀, et des variations spécifiques dans l'expression de certains micro-ARN plasmatiques. Ces micro-ARN, principalement impliqués dans les voies inflammatoires, révèlent des effets potentiels sur les mécanismes de régulation génétique et inflammatoire (26,32).

Le miR-223-3p est largement étudié pour son rôle dans la modulation de l'inflammation. Son association négative avec le NO₂ suggère que l'exposition à long terme à ce polluant pourrait altérer ses niveaux et, par conséquent, influencer les réponses inflammatoires. Cette observation est en accord avec certaines études qui ont montré une variation de l'expression de miR-223-3p en réponse à divers stimuli inflammatoires (58,59). Toutefois, des résultats contradictoires existent dans la littérature, ce qui reflète la complexité des interactions entre micro-ARN et environnement.

De même, le miR-146a-5p, connu pour sa régulation des voies inflammatoires via l'inhibiteur de NF- κ B, présente des résultats variés selon les études. Nos résultats montrent une association négative avec le NO₂, suggérant un potentiel effet modulateur de la pollution sur les réponses immunitaires (31,37,70,71). Cependant, des études antérieures ont rapporté des associations positives entre miR-146a-5p et pollution, soulignant la complexité des interactions entre micro-ARN et environnement (38).

Le miR-28-3p n'a pas été largement étudié dans le contexte de la pollution atmosphérique, mais nos résultats suggèrent une association négative. Cela pourrait refléter un effet direct ou indirect du NO₂ sur les processus de régulation génétique (74,101).

Le miR-23a-5p, malgré son rôle significatif dans la régulation des processus inflammatoires et fibrogéniques, a montré une association négative dans notre étude (72). Cette découverte est intéressante car elle pourrait indiquer des mécanismes de réponse cellulaire complexes face à l'exposition prolongée aux polluants.

L'association positive entre miR-451a et le NO₂ pourrait suggérer un mécanisme compensatoire ou adaptatif en réponse au stress oxydatif induit par la pollution (76–78). Le miR-451a est impliqué dans le transport de l'oxygène et la régulation du métabolisme cellulaire (75), et son élévation pourrait refléter des ajustements physiologiques face à une exposition accrue aux polluants. Cependant, des études antérieures, telles que celles de Motta et al. (79) et Deng et al. (81), ont montré des résultats variables sur l'association entre miR-451a et les niveaux de PM, ce qui souligne la nécessité de recherches supplémentaires.

II.Perspectives

Les micro-ARN jouent un rôle central dans la régulation épigénétique, influençant l'expression de nombreux gènes et divers processus biologiques et pathologiques (102,103). Leur identification comme biomarqueurs potentiels pour évaluer les effets des expositions environnementales sur la santé humaine s'inscrit dans une tendance observée dans la littérature actuelle, où ils sont de plus en plus reconnus comme des médiateurs des effets nocifs des polluants atmosphériques (25,70). Des études antérieures, bien que souvent réalisées sur des échantillons plus

réduits et avec des résultats parfois contradictoires, ont déjà mis en évidence des associations similaires (71,78). Ces variations peuvent s'expliquer par des différences dans les méthodes d'exposition, les populations étudiées, ou les types de polluants examinés.

La régulation des voies inflammatoires par les micro-ARN, notamment ceux identifiés dans notre étude, renforce l'idée que l'inflammation est un mécanisme clé par lequel la pollution atmosphérique exerce ses effets délétères sur la santé (72,104). Cette inflammation, lorsqu'elle est chronique ou mal régulée, est liée à une multitude de maladies, allant des troubles respiratoires comme la BPCO et l'asthme, aux maladies cardiovasculaires et aux cancers (37,58). Cependant, la corrélation observée entre certains micro-ARN de ménage, tels que miR-93-5p et miR-191-5p, et l'exposition à la pollution suggère des interactions complexes qui mériteraient d'être explorées plus en profondeur (74,79). Bien que les données sur l'exposition aient été modélisées avec une précision spatiale fine, d'autres facteurs environnementaux ou de style de vie non mesurés pourraient également influencer l'expression des micro-ARN (32,81).

Conclusion

En conclusion, cette étude suggère l'utilité des micro-ARN pour clarifier les relations complexes entre pollution atmosphérique et mécanismes physiopathologiques , en particulier les voies de l'inflammation. Cette voie de recherche ouvre des perspectives prometteuses pour le développement de biomarqueurs de risque environnemental, qui pourraient à terme contribuer à une meilleure prévention et gestion des maladies liées à la pollution.

Références

1. Principaux repères sur la qualité de l'air [Internet]. [cité 18 août 2024]. Disponible sur: [https://www.who.int/fr/news-room/fact-sheets/detail/ambient-\(outdoor\)-air-quality-and-health](https://www.who.int/fr/news-room/fact-sheets/detail/ambient-(outdoor)-air-quality-and-health)
2. Air pollution [Internet]. [cité 20 août 2024]. Disponible sur: <https://www.who.int/health-topics/air-pollution>
3. Strobl K, Irfan SA, Masood H, Latif N, Kurmi O. Association between PM10 exposure and risk of myocardial infarction in adults: A systematic review and meta-analysis. PLOS ONE [Internet]. 2024 [cité 20 août 2024];19(5). Disponible sur: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC11062553/>
4. Manisalidis I, Stavropoulou E, Stavropoulos A, Bezirtzoglou E. Environmental and Health Impacts of Air Pollution: A Review. Front Public Health. 20 févr 2020;8:14.
5. Particulate Matter (PM) Basics | US EPA [Internet]. [cité 18 août 2024]. Disponible sur: <https://www.epa.gov/pm-pollution/particulate-matter-pm-basics#PM>
6. Hamanaka RB, Mutlu GM. Particulate Matter Air Pollution: Effects on the Cardiovascular System. Front Endocrinol [Internet]. 2018 [cité 20 août 2024];9. Disponible sur: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6250783/>
7. Zhang L, Lu Q, Chang C. Epigenetics in Health and Disease. In: Chang C, Lu Q, éditeurs. Epigenetics in Allergy and Autoimmunity [Internet]. Singapore: Springer Singapore; 2020 [cité 18 août 2024]. p. 3-55. (Advances in Experimental Medicine and Biology; vol. 1253). Disponible sur: http://link.springer.com/10.1007/978-981-15-3449-2_1
8. Hartmann C, Corre-Menguy F, Boualem A, Jovanovic M, Lelandais-Brière C. Les microARN: Une nouvelle classe de régulateurs de l'expression génique. médecine/sciences. oct 2004;20(10):894-8.
9. Bushati N, Cohen SM. microRNA Functions. Annu Rev Cell Dev Biol. 1 nov 2007;23(Volume 23, 2007):175-205.
10. Searles CD. MicroRNAs and Cardiovascular Disease Risk. Curr Cardiol Rep. févr 2024;26(2):51-60.
11. The influence of epigenetics and inflammation on cardiometabolic risks. Semin Cell Dev Biol. 15 févr 2024;154:175-84.
12. Brook RD, Rajagopalan S, C. Arden Pope III, Brook JR, Bhatnagar A, Diez-Roux AV, et al. Particulate Matter Air Pollution and Cardiovascular Disease. Circulation [Internet]. 1 juin 2010 [cité 19 août 2024]; Disponible sur: <https://www.ahajournals.org/doi/10.1161/CIR.0b013e3181d8e1>
13. Humphries B, Wang Z, Yang C. MicroRNA Regulation of Epigenetic Modifiers in Breast Cancer. Cancers. juill 2019;11(7):897.
14. Khordadmehr M, Jigari-Asl F, Ezzati H, Shahbazi R, Sadreddini S, Safaei S, et al. A comprehensive review on miR-451: A promising cancer biomarker with therapeutic potential. J Cell Physiol. déc 2019;234(12):21716-31.
15. Huang Z, Zhang L, Zhu D, Shan X, Zhou X, Qi LW, et al. A novel serum microRNA signature to screen esophageal squamous cell carcinoma. Cancer Med. janv 2017;6(1):109-19.

16. Riant M, Meirhaeghe A, Giovannelli J, Occelli F, Havet A, Cuny D, et al. Associations between long-term exposure to air pollution, glycosylated hemoglobin, fasting blood glucose and diabetes mellitus in northern France. *Environ Int.* 1 nov 2018;120:121-9.
17. Darras-Hostens M, Achour D, Muntaner M, Grare C, Zarcone G, Garçon G, et al. Short-term and residential exposure to air pollution: Associations with inflammatory biomarker levels in adults living in northern France. *Sci Total Environ.* août 2022;833:154985.
18. Landgraf P, Rusu M, Sheridan R, Sewer A, Iovino N, Aravin A, et al. A Mammalian microRNA Expression Atlas Based on Small RNA Library Sequencing. *Cell.* 29 juin 2007;129(7):1401-14.
19. Pozniak T, Shcharbin D, Bryszewska M. Circulating microRNAs in Medicine. *Int J Mol Sci* [Internet]. avr 2022 [cité 19 août 2024];23(7). Disponible sur: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8999557/>
20. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, Pogosova-Agadjanyan EL, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci.* 29 juill 2008;105(30):10513-8.
21. Gilad S, Meiri E, Yogev Y, Benjamin S, Lebanony D, Yerushalmi N, et al. Serum MicroRNAs Are Promising Novel Biomarkers. Williams S, éditeur. *PLoS ONE.* 5 sept 2008;3(9):e3148.
22. Fujimoto S, Manabe S, Morimoto C, Ozeki M, Hamano Y, Hirai E, et al. Distinct spectrum of microRNA expression in forensically relevant body fluids and probabilistic discriminant approach. *Sci Rep.* 4 oct 2019;9(1):1-10.
23. Hunter MP, Ismail N, Zhang X, Aguda BD, Lee EJ, Yu L, et al. Detection of microRNA Expression in Human Peripheral Blood Microvesicles. Lo YMD, éditeur. *PLoS ONE.* 11 nov 2008;3(11):e3694.
24. Krauskopf J, de Kok TM, Hebels DG, Bergdahl IA, Johansson A, Spaeth F, et al. MicroRNA profile for health risk assessment: Environmental exposure to persistent organic pollutants strongly affects the human blood microRNA machinery. *Sci Rep.* 23 août 2017;7(1):1-9.
25. Krauskopf J, Caiment F, Van Veldhoven K, Chadeau-Hyam M, Sinharay R, Chung KF, et al. The human circulating miRNome reflects multiple organ disease risks in association with short-term exposure to traffic-related air pollution. *Environ Int.* avr 2018;113:26-34.
26. Rodosthenous RS, Coull BA, Lu Q, Vokonas PS, Schwartz JD, Baccarelli AA. Ambient particulate matter and microRNAs in extracellular vesicles: a pilot study of older individuals. *Part Fibre Toxicol.* 8 mars 2016;13:13.
27. Rodosthenous RS, Kloog I, Colicino E, Zhong J, Herrera LA, Vokonas P, et al. Extracellular vesicle-enriched microRNAs interact in the association between long-term particulate matter and blood pressure in elderly men. *Environ Res.* 1 nov 2018;167:640-9.
28. Cheng M, Wang B, Yang M, Ma J, Ye Z, Xie L, et al. microRNAs expression in relation to particulate matter exposure: A systematic review. *Environ Pollut.* mai 2020;260:113961.
29. Krauskopf J, Verheijen M, Kleinjans JC, Kok TM de, Caiment F. Development and Regulatory Application of MicroRNA Biomarkers. *Biomark Med* [Internet]. 1 nov 2015 [cité 19 août 2024]; Disponible sur: <https://www.tandfonline.com/doi/abs/10.2217/bmm.15.50>

30. Chen H, Zhang S, Yu B, Xu Y, Rappold AG, Diaz-Sanchez D, et al. Circulating microRNAs as putative mediators in the association between short-term exposure to ambient air pollution and cardiovascular biomarkers. *Ecotoxicol Environ Saf.* juill 2022;239:113604.
31. Fossati S, Baccarelli A, Zanobetti A, Hoxha M, Vokonas PS, Wright RO, et al. Ambient particulate air pollution and microRNAs in elderly men. *Epidemiol Camb Mass.* janv 2014;25(1):68-78.
32. Li J, Wang T, Wang Y, Xu M, Zhang L, Li X, et al. Particulate matter air pollution and the expression of microRNAs and pro-inflammatory genes: Association and mediation among children in Jinan, China. *J Hazard Mater.* mai 2020;389:121843.
33. Quach A, Giovannelli J, Chérot-Kornobis N, Ciuchete A, Clément G, Matran R, et al. Prevalence and underdiagnosis of airway obstruction among middle-aged adults in northern France: The ELISABET study 2011–2013. *Respir Med.* 1 déc 2015;109(12):1553-61.
34. Dauchet L, Hulo S, Cherot-Kornobis N, Matran R, Amouyel P, Edmé JL, et al. Short-term exposure to air pollution: Associations with lung function and inflammatory markers in non-smoking, healthy adults. *Environ Int.* déc 2018;121:610-9.
35. Havet A, Hulo S, Cuny D, Riant M, Occelli F, Cherot-Kornobis N, et al. Residential exposure to outdoor air pollution and adult lung function, with focus on small airway obstruction. *Environ Res.* avr 2020;183:109161.
36. BT2015_MEL_VF.pdf [Internet]. [cité 19 août 2024]. Disponible sur: https://www.atmo-hdf.fr/sites/hdf/files/content/migrated/Bilans-territoriaux/BT2015_MEL_VF.pdf
37. Chen R, Li H, Cai J, Wang C, Lin Z, Liu C, et al. Fine Particulate Air Pollution and the Expression of microRNAs and Circulating Cytokines Relevant to Inflammation, Coagulation, and Vasoconstriction. *Environ Health Perspect.* 17 janv 2018;126(1):017007.
38. Krauskopf J, van Veldhoven K, Chadeau-Hyam M, Vermeulen R, Carrasco-Turigas G, Nieuwenhuijsen M, et al. Short-term exposure to traffic-related air pollution reveals a compound-specific circulating miRNA profile indicating multiple disease risks. *Environ Int.* juill 2019;128:193-200.
39. Louwies T, Vuegen C, Panis LI, Cox B, Vrijens K, Nawrot TS, et al. miRNA expression profiles and retinal blood vessel calibers are associated with short-term particulate matter air pollution exposure. *Environ Res.* mai 2016;147:24-31.
40. Quezada-Maldonado EM, Sánchez-Pérez Y, Chirino YI, Vaca-Paniagua F, García-Cuellar CM. miRNAs deregulation in lung cells exposed to airborne particulate matter (PM10) is associated with pathways deregulated in lung tumors. *Environ Pollut.* 1 oct 2018;241:351-8.
41. Ruiz-Vera T, Ochoa-Martínez AC, Pruneda-Álvarez LG, Zarazúa S, Pérez-Maldonado IN. Exposure to biomass smoke is associated with an increased expression of circulating miRNA-126 and miRNA-155 in Mexican women: a pilot study. *Drug Chem Toxicol* [Internet]. 4 mai 2019 [cité 19 août 2024]; Disponible sur: <https://www.tandfonline.com/doi/abs/10.1080/01480545.2018.1526181>
42. Fry RC, Rager JE, Bauer R, Sebastian E, Peden DB, Jaspers I, et al. Air toxics and epigenetic effects: ozone altered microRNAs in the sputum of human subjects. *Am J Physiol Lung Cell Mol Physiol.* 15 juin 2014;306(12):L1129-1137.
43. Sima M, Rossnerova A, Simova Z, Rossner P. The Impact of Air Pollution Exposure on the MicroRNA Machinery and Lung Cancer Development. *J Pers Med.* janv 2021;11(1):60.

44. Ebrahimi A, Sadroddiny E. MicroRNAs in lung diseases: Recent findings and their pathophysiological implications. *Pulm Pharmacol Ther.* oct 2015;34:55-63.
45. Rupani H, Sanchez-Elsner T, Howarth P. MicroRNAs and respiratory diseases. *Eur Respir J.* mars 2013;41(3):695-705.
46. Sessa R, Hata A. Role of microRNAs in Lung Development and Pulmonary Diseases. *Pulm Circ [Internet].* 1 avr 2013 [cité 19 août 2024]; Disponible sur: <https://journals.sagepub.com/doi/full/10.4103/2045-8932.114758>
47. Alipoor SD, Adcock IM, Garssen J, Mortaz E, Varahram M, Mirsaedi M, et al. The roles of miRNAs as potential biomarkers in lung diseases. *Eur J Pharmacol.* nov 2016;791:395-404.
48. Moretti F, D'Antona P, Finardi E, Barbetta M, Dominioni L, Poli A, et al. Systematic review and critique of circulating miRNAs as biomarkers of stage I-II non-small cell lung cancer. *Oncotarget.* 11 nov 2017;8(55):94980.
49. Shah JS, Soon PS, Marsh DJ. Comparison of Methodologies to Detect Low Levels of Hemolysis in Serum for Accurate Assessment of Serum microRNAs. *PLOS ONE.* 7 avr 2016;11(4):e0153200.
50. Blondal T, Jensby Nielsen S, Baker A, Andreasen D, Mouritzen P, Wrang Teilum M, et al. Assessing sample and miRNA profile quality in serum and plasma or other biofluids. *Methods.* janv 2013;59(1):S1-6.
51. Donati S, Ciuffi S, Brandi ML. Human Circulating miRNAs Real-time qRT-PCR-based Analysis: An Overview of Endogenous Reference Genes Used for Data Normalization. *Int J Mol Sci.* janv 2019;20(18):4353.
52. Andersen CL, Jensen JL, Ørntoft TF. Normalization of Real-Time Quantitative Reverse Transcription-PCR Data: A Model-Based Variance Estimation Approach to Identify Genes Suited for Normalization, Applied to Bladder and Colon Cancer Data Sets. *Cancer Res.* 1 août 2004;64(15):5245-50.
53. Heredia FP de, Gómez-Martínez S, Marcos A. Obesity, inflammation and the immune system. *Proc Nutr Soc.* mai 2012;71(2):332-8.
54. Khan J, Lieberman JA, Lockwood CM. Variability in, variability out: best practice recommendations to standardize pre-analytical variables in the detection of circulating and tissue microRNAs. *Clin Chem Lab Med CCLM.* 1 mai 2017;55(5):608-21.
55. Broz P, Dixit VM. Inflammasomes: mechanism of assembly, regulation and signalling. *Nat Rev Immunol.* juill 2016;16(7):407-20.
56. Bauernfeind FG, Horvath G, Stutz A, Alnemri ES, MacDonald K, Speert D, et al. Cutting edge: NF-kappaB activating pattern recognition and cytokine receptors license NLRP3 inflammasome activation by regulating NLRP3 expression. *J Immunol Baltim Md 1950.* 15 juill 2009;183(2):787-91.
57. Olivieri F, Prattichizzo F, Giuliani A, Matacchione G, Rippo MR, Sabbatinelli J, et al. miR-21 and miR-146a: The microRNAs of inflammaging and age-related diseases. *Ageing Res Rev.* sept 2021;70:101374.
58. Bauernfeind F, Rieger A, Schildberg FA, Knolle PA, Schmid-Burgk JL, Hornung V. NLRP3 inflammasome activity is negatively controlled by miR-223. *J Immunol Baltim Md 1950.* 15 oct 2012;189(8):4175-81.

59. Haneklaus M, Gerlic M, Kurowska-Stolarska M, Rainey AA, Pich D, McInnes IB, et al. Cutting edge: miR-223 and EBV miR-BART15 regulate the NLRP3 inflammasome and IL-1 β production. *J Immunol Baltim Md 1950*. 15 oct 2012;189(8):3795-9.
60. Herberth G, Bauer M, Gasch M, Hinz D, Röder S, Olek S, et al. Maternal and cord blood miR-223 expression associates with prenatal tobacco smoke exposure and low regulatory T-cell numbers. *J Allergy Clin Immunol*. févr 2014;133(2):543-50.
61. Schembri F, Sridhar S, Perdomo C, Gustafson AM, Zhang X, Ergun A, et al. MicroRNAs as modulators of smoking-induced gene expression changes in human airway epithelium. *Proc Natl Acad Sci U S A*. 17 févr 2009;106(7):2319-24.
62. Izzotti A, Calin GA, Arrigo P, Steele VE, Croce CM, De Flora S. Downregulation of microRNA expression in the lungs of rats exposed to cigarette smoke. *FASEB J Off Publ Fed Am Soc Exp Biol*. mars 2009;23(3):806-12.
63. Carberry CK, Koval LE, Payton A, Hartwell H, Ho Kim Y, Smith GJ, et al. Wildfires and extracellular vesicles: Exosomal MicroRNAs as mediators of cross-tissue cardiopulmonary responses to biomass smoke. *Environ Int*. sept 2022;167:107419.
64. Hou J, Deng Q, Deng X, Zhong W, Liu S, Zhong Z. MicroRNA-146a-5p alleviates lipopolysaccharide-induced NLRP3 inflammasome injury and pro-inflammatory cytokine production via the regulation of TRAF6 and IRAK1 in human umbilical vein endothelial cells (HUVECs). *Ann Transl Med*. sept 2021;9(18):1433.
65. Etzrodt M, Cortez-Retamozo V, Newton A, Zhao J, Ng A, Wildgruber M, et al. Regulation of monocyte functional heterogeneity by miR-146a and Relb. *Cell Rep*. 19 avr 2012;1(4):317-24.
66. McMillan DH, Woeller CF, Thatcher TH, Spinelli SL, Maggirwar SB, Sime PJ, et al. Attenuation of inflammatory mediator production by the NF- κ B member RelB is mediated by microRNA-146a in lung fibroblasts. *Am J Physiol Lung Cell Mol Physiol*. 1 juin 2013;304(11):L774-781.
67. Boldin MP, Taganov KD, Rao DS, Yang L, Zhao JL, Kalwani M, et al. miR-146a is a significant brake on autoimmunity, myeloproliferation, and cancer in mice. *J Exp Med*. 6 juin 2011;208(6):1189-201.
68. Zhao JL, Rao DS, Boldin MP, Taganov KD, O'Connell RM, Baltimore D. NF-kappaB dysregulation in microRNA-146a-deficient mice drives the development of myeloid malignancies. *Proc Natl Acad Sci U S A*. 31 mai 2011;108(22):9184-9.
69. Bollati V, Marinelli B, Apostoli P, Bonzini M, Nordio F, Hoxha M, et al. Exposure to metal-rich particulate matter modifies the expression of candidate microRNAs in peripheral blood leukocytes. *Environ Health Perspect*. juin 2010;118(6):763-8.
70. Tsamou M, Vrijens K, Madhloum N, Lefebvre W, Vanpoucke C, Nawrot TS. Air pollution-induced placental epigenetic alterations in early life: a candidate miRNA approach. *Epigenetics*. 2018;13(2):135-46.
71. Chang H, Chang H, Cheng T, Lee GD, Chen X, Qi K. Micro-ribonucleic acid-23a-3p prevents the onset of type 2 diabetes mellitus by suppressing the activation of nucleotide-binding oligomerization-like receptor family pyrin domain containing 3 inflammatory bodies-caused pyroptosis through negatively regulating NIMA-related kinase 7. *J Diabetes Investig*. mars 2021;12(3):334-45.

72. Pan Z, Shan Q, Gu P, Wang XM, Tai LW, Sun M, et al. miRNA-23a/CXCR4 regulates neuropathic pain via directly targeting TXNIP/NLRP3 inflammasome axis. *J Neuroinflammation*. 31 janv 2018;15(1):29.
73. Yang M, Yao Y, Eades G, Zhang Y, Zhou Q. MiR-28 regulates Nrf2 expression through a Keap1-independent mechanism. *Breast Cancer Res Treat*. oct 2011;129(3):983-91.
74. Ichimura Y, Waguri S, Sou YS, Kageyama S, Hasegawa J, Ishimura R, et al. Phosphorylation of p62 activates the Keap1-Nrf2 pathway during selective autophagy. *Mol Cell*. 12 sept 2013;51(5):618-31.
75. Hur W, Lee JH, Kim SW, Kim JH, Bae SH, Kim M, et al. Downregulation of microRNA-451 in non-alcoholic steatohepatitis inhibits fatty acid-induced proinflammatory cytokine production through the AMPK/AKT pathway. *Int J Biochem Cell Biol*. juill 2015;64:265-76.
76. Li HP, Zeng XC, Zhang B, Long JT, Zhou B, Tan GS, et al. miR-451 inhibits cell proliferation in human hepatocellular carcinoma through direct suppression of IKK- β . *Carcinogenesis*. nov 2013;34(11):2443-51.
77. Xie J, Hu X, Yi C, Hu G, Zhou X, Jiang H. MicroRNA-451 protects against cardiomyocyte anoxia/reoxygenation injury by inhibiting high mobility group box 1 expression. *Mol Med Rep*. juin 2016;13(6):5335-41.
78. Barnay-Verdier S, Maréchal V, Borde C. [HMGB1: a link between innate and adaptive immunity]. *Rev Francoph Lab RFL*. déc 2009;2009(417):59-68.
79. Motta V, Favero C, Dioni L, Iodice S, Battaglia C, Angelici L, et al. MicroRNAs are associated with blood-pressure effects of exposure to particulate matter: Results from a mediated moderation analysis. *Environ Res*. avr 2016;146:274-81.
80. Dubowsky SD, Suh H, Schwartz J, Coull BA, Gold DR. Diabetes, obesity, and hypertension may enhance associations between air pollution and markers of systemic inflammation. *Environ Health Perspect*. juill 2006;114(7):992-8.
81. Deng Q, Dai X, Feng W, Huang S, Yuan Y, Xiao Y, et al. Co-exposure to metals and polycyclic aromatic hydrocarbons, microRNA expression, and early health damage in coke oven workers. *Environ Int*. janv 2019;122:369-80.
82. Boxberger N, Hecker M, Zettl UK. Dysregulation of Inflammasome Priming and Activation by MicroRNAs in Human Immune-Mediated Diseases. *J Immunol Baltim Md 1950*. 15 avr 2019;202(8):2177-87.
83. Mortazavi-Jahromi SS, Aslani M, Mirshafiey A. A comprehensive review on miR-146a molecular mechanisms in a wide spectrum of immune and non-immune inflammatory diseases. *Immunol Lett*. nov 2020;227:8-27.
84. Roffel MP, Bracke KR, Heijink IH, Maes T. miR-223: A Key Regulator in the Innate Immune Response in Asthma and COPD. *Front Med*. 19 mai 2020;7:527739.
85. Specjalski K, Jassem E. MicroRNAs: Potential Biomarkers and Targets of Therapy in Allergic Diseases? *Arch Immunol Ther Exp (Warsz)*. août 2019;67(4):213-23.

86. Aslani M, Mortazavi-Jahromi SS, Mirshafiey A. Efficient roles of miR-146a in cellular and molecular mechanisms of neuroinflammatory disorders: An effectual review in neuroimmunology. *Immunol Lett.* oct 2021;238:1-20.
87. Vickers KC, Landstreet SR, Levin MG, Shoucri BM, Toth CL, Taylor RC, et al. MicroRNA-223 coordinates cholesterol homeostasis. *Proc Natl Acad Sci U S A.* 7 oct 2014;111(40):14518-23.
88. Grieco GE, Besharat ZM, Licata G, Fignani D, Brusco N, Nigi L, et al. Circulating microRNAs as clinically useful biomarkers for Type 2 Diabetes Mellitus: miRNomics from bench to bedside. *Transl Res J Lab Clin Med.* sept 2022;247:137-57.
89. Kaur P, Kotru S, Singh S, Munshi A. Role of miRNAs in diabetic neuropathy: mechanisms and possible interventions. *Mol Neurobiol.* mars 2022;59(3):1836-49.
90. Iacona JR, Lutz CS. miR-146a-5p: Expression, regulation, and functions in cancer. *Wiley Interdiscip Rev RNA.* juill 2019;10(4):e1533.
91. Favero A, Segatto I, Perin T, Belletti B. The many facets of miR-223 in cancer: Oncosuppressor, oncogenic driver, therapeutic target, and biomarker of response. *Wiley Interdiscip Rev RNA.* nov 2021;12(6):e1659.
92. Wang N, Tan HY, Feng YG, Zhang C, Chen F, Feng Y. microRNA-23a in Human Cancer: Its Roles, Mechanisms and Therapeutic Relevance. *Cancers.* 20 déc 2018;11(1):E7.
93. Silva CMS, Barros-Filho MC, Wong DVT, Mello JBH, Nobre LMS, Wanderley CWS, et al. Circulating let-7e-5p, miR-106a-5p, miR-28-3p, and miR-542-5p as a Promising microRNA Signature for the Detection of Colorectal Cancer. *Cancers.* 24 mars 2021;13(7):1493.
94. Ghafouri-Fard S, Abak A, Talebi SF, Shoorei H, Branicki W, Taheri M, et al. Role of miRNA and lncRNAs in organ fibrosis and aging. *Biomed Pharmacother Biomedecine Pharmacother.* nov 2021;143:112132.
95. Wang B, Duan X, Xu Q, Li Y. Diagnostic and prognostic significance of miR-451a in patients with atherosclerosis. *Vascular.* 17 déc 2021;17085381211058571.
96. Churov AV, Oleinik EK, Knip M. MicroRNAs in rheumatoid arthritis: altered expression and diagnostic potential. *Autoimmun Rev.* nov 2015;14(11):1029-37.
97. Kyyaly MA, Sanchez-Elsner T, He P, Sones CL, Arshad SH, Kurukulaaratchy RJ. Circulating miRNAs-A potential tool to identify severe asthma risk? *Clin Transl Allergy.* juin 2021;11(4):e12040.
98. Akbas F, Coskunpinar E, Aynaci E, Oltulu YM, Yildiz P. Analysis of serum micro-RNAs as potential biomarker in chronic obstructive pulmonary disease. *Exp Lung Res.* août 2012;38(6):286-94.
99. Du X, Zhang Q, Jiang Y, Zhu X, Zhang Y, Liu C, et al. Characterization of plasma-derived exosomal miRNA changes following traffic-related air pollution exposure: A randomized, crossover trial based on small RNA sequencing. *Environ Int.* sept 2022;167:107430.
100. Bohatá J, Horváthová V, Pavlíková M, Stibůrková B. Circulating microRNA alternations in primary hyperuricemia and gout. *Arthritis Res Ther.* déc 2021;23(1):186.
101. Jain A, Lamark T, Sjøttem E, Larsen KB, Awuh JA, Øvervatn A, et al. p62/SQSTM1 is a target gene for transcription factor NRF2 and creates a positive feedback loop by inducing antioxidant response element-driven gene transcription. *J Biol Chem.* 16 juill 2010;285(29):22576-91.

102. Bartel DP. MicroRNAs: Genomics, Biogenesis, Mechanism, and Function. *Cell*. 23 janv 2004;116(2):281-97.
103. He L, He X, Lim LP, de Stanchina E, Xuan Z, Liang Y, et al. A microRNA component of the p53 tumour suppressor network. *Nature*. 28 juin 2007;447(7148):1130-4.
104. Jin W, Fei X, Wang X, Chen F, Song Y. Circulating miRNAs as Biomarkers for Prostate Cancer Diagnosis in Subjects with Benign Prostatic Hyperplasia. *J Immunol Res*. 8 mai 2020;2020:5873056.
105. Tsamou M, Vrijens K, Madhloum N, Lefebvre W, Vanpoucke C, Nawrot TS. Air pollution-induced placental epigenetic alterations in early life: a candidate miRNA approach. *Epigenetics*. 2018;13(2):135-46.
106. Rider CF, Yamamoto M, Günther OP, Hirota JA, Singh A, Tebbutt SJ, et al. Controlled diesel exhaust and allergen coexposure modulates microRNA and gene expression in humans: Effects on inflammatory lung markers. *J Allergy Clin Immunol*. déc 2016;138(6):1690-700.
107. Du X, Zhang Q, Jiang Y, Zhu X, Zhang Y, Liu C, et al. Characterization of plasma-derived exosomal miRNA changes following traffic-related air pollution exposure: A randomized, crossover trial based on small RNA sequencing. *Environ Int*. sept 2022;167:107430.
108. Chen R, Li H, Cai J, Wang C, Lin Z, Liu C, et al. Fine Particulate Air Pollution and the Expression of microRNAs and Circulating Cytokines Relevant to Inflammation, Coagulation, and Vasoconstriction. *Environ Health Perspect*. 17 janv 2018;126(1):017007.
109. Krauskopf J, Caiment F, van Veldhoven K, Chadeau-Hyam M, Sinharay R, Chung KF, et al. The human circulating miRNome reflects multiple organ disease risks in association with short-term exposure to traffic-related air pollution. *Environ Int*. avr 2018;113:26-34.
110. Krauskopf J, van Veldhoven K, Chadeau-Hyam M, Vermeulen R, Carrasco-Turigas G, Nieuwenhuijsen M, et al. Short-term exposure to traffic-related air pollution reveals a compound-specific circulating miRNA profile indicating multiple disease risks. *Environ Int*. juill 2019;128:193-200.
111. Hou L, Barupal J, Zhang W, Zheng Y, Liu L, Zhang X, et al. Particulate Air Pollution Exposure and Expression of Viral and Human MicroRNAs in Blood: The Beijing Truck Driver Air Pollution Study. *Environ Health Perspect*. mars 2016;124(3):344-50.
112. Vriens A, Nawrot TS, Saenen ND, Provost EB, Kicinski M, Lefebvre W, et al. Recent exposure to ultrafine particles in school children alters miR-222 expression in the extracellular fraction of saliva. *Environ Health Glob Access Sci Source*. 26 juill 2016;15(1):80.
113. Mancini FR, Laine JE, Tarallo S, Vlaanderen J, Vermeulen R, van Nunen E, et al. microRNA expression profiles and personal monitoring of exposure to particulate matter. *Environ Pollut Barking Essex* 1987. août 2020;263(Pt B):114392.
114. Chen H, Xu Y, Rappold A, Diaz-Sanchez D, Tong H. Effects of ambient ozone exposure on circulating extracellular vehicle microRNA levels in coronary artery disease patients. *J Toxicol Environ Health A*. 2 mai 2020;83(9):351-62.
115. Louwies T, Vuegen C, Panis LI, Cox B, Vrijens K, Nawrot TS, et al. miRNA expression profiles and retinal blood vessel calibers are associated with short-term particulate matter air pollution exposure. *Environ Res*. mai 2016;147:24-31.

116. Rodosthenous RS, Coull BA, Lu Q, Vokonas PS, Schwartz JD, Baccarelli AA. Ambient particulate matter and microRNAs in extracellular vesicles: a pilot study of older individuals. *Part Fibre Toxicol.* 8 mars 2016;13:13.
117. Fossati S, Baccarelli A, Zanobetti A, Hoxha M, Vokonas PS, Wright RO, et al. Ambient particulate air pollution and microRNAs in elderly men. *Epidemiol Camb Mass.* janv 2014;25(1):68-78.
118. Motta V, Favero C, Dioni L, Iodice S, Battaglia C, Angelici L, et al. MicroRNAs are associated with blood-pressure effects of exposure to particulate matter: Results from a mediated moderation analysis. *Environ Res.* avr 2016;146:274-81.
119. Li J, Wang T, Wang Y, Xu M, Zhang L, Li X, et al. Particulate matter air pollution and the expression of microRNAs and pro-inflammatory genes: Association and mediation among children in Jinan, China. *J Hazard Mater.* 5 mai 2020;389:121843.
120. Matsushashi S, Manirujjaman M, Hamajima H, Ozaki I. Control Mechanisms of the Tumor Suppressor PDCD4: Expression and Functions. *Int J Mol Sci.* 9 mai 2019;20(9):2304.
121. Xue Z, Xi Q, Liu H, Guo X, Zhang J, Zhang Z, et al. miR-21 promotes NLRP3 inflammasome activation to mediate pyroptosis and endotoxic shock. *Cell Death Dis.* 12 juin 2019;10(6):461.
122. Chang H, Chang H, Cheng T, Lee GD, Chen X, Qi K. Micro-ribonucleic acid-23a-3p prevents the onset of type 2 diabetes mellitus by suppressing the activation of nucleotide-binding oligomerization-like receptor family pyrin domain containing 3 inflammatory bodies-caused pyroptosis through negatively regulating NIMA-related kinase 7. *J Diabetes Investig.* mars 2021;12(3):334-45.
123. Pan Z, Shan Q, Gu P, Wang XM, Tai LW, Sun M, et al. miRNA-23a/CXCR4 regulates neuropathic pain via directly targeting TXNIP/NLRP3 inflammasome axis. *J Neuroinflammation.* 31 janv 2018;15(1):29.
124. Yang M, Yao Y, Eades G, Zhang Y, Zhou Q. MiR-28 regulates Nrf2 expression through a Keap1-independent mechanism. *Breast Cancer Res Treat.* oct 2011;129(3):983-91.
125. Zhou T, Chen Y li. The Functional Mechanisms of miR-30b-5p in Acute Lung Injury in Children. *Med Sci Monit.* 2 janv 2019;25:40-51.
126. Chen Q, Hu H, Jiao D, Yan J, Xu W, Tang X, et al. miR-126-3p and miR-451a correlate with clinicopathological features of lung adenocarcinoma: The underlying molecular mechanisms. *Oncol Rep.* août 2016;36(2):909-17.
127. Guo C, Sah JF, Beard L, Willson JKV, Markowitz SD, Guda K. The noncoding RNA, miR-126, suppresses the growth of neoplastic cells by targeting phosphatidylinositol 3-kinase signaling and is frequently lost in colon cancers. *Genes Chromosomes Cancer.* nov 2008;47(11):939-46.
128. Zhang J, Du Y ying, Lin Y feng, Chen Y ting, Yang L, Wang H jun, et al. The cell growth suppressor, mir-126, targets IRS-1. *Biochem Biophys Res Commun.* déc 2008;377(1):136-40.
129. Sibilano M, Tullio V, Adorno G, Savini I, Gasperi V, Catani MV. Platelet-Derived miR-126-3p Directly Targets AKT2 and Exerts Anti-Tumor Effects in Breast Cancer Cells: Further Insights in Platelet-Cancer Interplay. *Int J Mol Sci.* 13 mai 2022;23(10):5484.
130. Han S, Lin F, Ruan Y, Zhao S, Yuan R, Ning J, et al. miR-132-3p promotes the cisplatin-induced apoptosis and inflammatory response of renal tubular epithelial cells by targeting SIRT1 via the NF- κ B pathway. *Int Immunopharmacol.* oct 2021;99:108022.

131. Olivieri F, Prattichizzo F, Giuliani A, Matacchione G, Rippo MR, Sabbatinelli J, et al. miR-21 and miR-146a: The microRNAs of inflammaging and age-related diseases. *Ageing Res Rev.* sept 2021;70:101374.
132. Hou J, Deng Q, Deng X, Zhong W, Liu S, Zhong Z. MicroRNA-146a-5p alleviates lipopolysaccharide-induced NLRP3 inflammasome injury and pro-inflammatory cytokine production via the regulation of TRAF6 and IRAK1 in human umbilical vein endothelial cells (HUVECs). *Ann Transl Med.* sept 2021;9(18):1433.
133. Department of Gastroenterology, The Affiliated Jiangning Hospital of Nanjing Medical University, Jiangsu, China, Xia F, Bo W, Department of Gastroenterology, The Affiliated Jiangning Hospital of Nanjing Medical University, Jiangsu, China, Ding J, Department of Gastroenterology, The Affiliated Jiangning Hospital of Nanjing Medical University, Jiangsu, China, et al. MiR-222-3p Aggravates the Inflammatory Response by Targeting SOCS1 to Activate STAT3 Signaling in Ulcerative Colitis. *Turk J Gastroenterol.* 21 nov 2022;33(11):934-44.
134. Zhang P, Yu J, Gui Y, Sun C, Han W. Inhibition of miRNA-222-3p Relieves Staphylococcal Enterotoxin B-Induced Liver Inflammatory Injury by Upregulating Suppressors of Cytokine Signaling 1. *Yonsei Med J.* 2019;60(11):1093.
135. Bauernfeind F, Rieger A, Schildberg FA, Knolle PA, Schmid-Burgk JL, Hornung V. NLRP3 inflammasome activity is negatively controlled by miR-223. *J Immunol Baltim Md 1950.* 15 oct 2012;189(8):4175-81.
136. Haneklaus M, Gerlic M, Kurowska-Stolarska M, Rainey AA, Pich D, McInnes IB, et al. Cutting edge: miR-223 and EBV miR-BART15 regulate the NLRP3 inflammasome and IL-1 β production. *J Immunol Baltim Md 1950.* 15 oct 2012;189(8):3795-9.
137. Hur W, Lee JH, Kim SW, Kim JH, Bae SH, Kim M, et al. Downregulation of microRNA-451 in non-alcoholic steatohepatitis inhibits fatty acid-induced proinflammatory cytokine production through the AMPK/AKT pathway. *Int J Biochem Cell Biol.* juill 2015;64:265-76.
138. Li HP, Zeng XC, Zhang B, Long JT, Zhou B, Tan GS, et al. miR-451 inhibits cell proliferation in human hepatocellular carcinoma through direct suppression of IKK- β . *Carcinogenesis.* nov 2013;34(11):2443-51.
139. Xie J, Hu X, Yi C, Hu G, Zhou X, Jiang H. MicroRNA-451 protects against cardiomyocyte anoxia/reoxygenation injury by inhibiting high mobility group box 1 expression. *Mol Med Rep.* juin 2016;13(6):5335-41.
140. Barnay-Verdier S, Maréchal V, Borde C. [HMGB1: a link between innate and adaptive immunity]. *Rev Francoph Lab RFL.* déc 2009;2009(417):59-68.
141. Fabbri E, Montagner G, Bianchi N, Finotti A, Borgatti M, Lampronti I, et al. MicroRNA miR-93-5p regulates expression of IL-8 and VEGF in neuroblastoma SK-N-AS cells. *Oncol Rep.* mai 2016;35(5):2866-72.
142. Zhang C, Nie P, Zhou C, Hu Y, Duan S, Gu M, et al. Oxidative stress-induced mitophagy is suppressed by the miR-106b-93-25 cluster in a protective manner. *Cell Death Dis.* 24 févr 2021;12(2):209.
143. Chen J, Sun J, Hu Y, Wan X, Wang Y, Gao M, et al. MicroRNA-191-5p ameliorates amyloid- β ₁₋₄₀-mediated retinal pigment epithelium cell injury by suppressing the NLRP3 inflammasome

pathway. *FASEB J* [Internet]. avr 2021 [cité 2 mars 2023];35(4). Disponible sur: <https://onlinelibrary.wiley.com/doi/10.1096/fj.202000645RR>

144. Wan W, Liu G, Li X, Liu Y, Wang Y, Pan H, et al. MiR-191-5p alleviates microglial cell injury by targeting Map3k12 (mitogen-activated protein kinase kinase kinase 12) to inhibit the MAPK (mitogen-activated protein kinase) signaling pathway in Alzheimer's disease. *Bioengineered*. 20 déc 2021;12(2):12678-90.

Annexes

Supplemental table 1 : Levels of miRNA

micro-RNA	Mean(SD)*
miR-21-5p	0.22 (0.648)
miR-23a-5p	0.573 (0.887)
miR-28-3p	-4.468 (0.779)
miR-30b-5p	-1.848 (0.874)
miR-126-3p	1.152 (0.604)
miR-132-3p	-5.434 (0.795)
miR-146a-5p	-0.952 (0.865)
miR-222-3p	-2.717 (0.703)
miR-223-3p	1.943 (0.644)
miR-451a	3.47 (1.158)
miR-93-5p	0.84 (0.58)
miR-191-5p	-0.84 (0.58)

* Expressed in difference of Ct between miRNA and average of the two housekeeping micro-ARN(miR-93-5p and miR-191-5p)

Supplemental table 2 : Association between residential PM₁₀ and miRNA using miRNA minus- average of the two housekeeping micro-ARN.

micro-RNA	men			women		
	PM ₁₀ (per 2µg/m ³ increase)			PM ₁₀ (per 2µg/m ³ increase)		
	% of variation [IC95%]	p	p FDR	% of variation [IC95%]	p	p FDR*
miR-21-5p	1.4 [-2.8 ; 5.8]	0.520	0.743	-0.3 [-4.9 ; 4.4]	0.884	0.982
miR-23a-5p	-2.8 [-8.4 ; 3.2]	0.360	0.721	-3.2 [-9 ; 3.0]	0.309	0.515
miR-28-3p	-5.9 [-10.7 ; -0.8]	0.024	0.237	-2.7 [-7.7 ; 2.6]	0.315	0.450
miR-30b-5p	1.4 [-4.1 ; 7.2]	0.619	0.773	-4.9 [-10.2 ; 0.8]	0.089	0.179
miR-126-3p	0.4 [-3.6 ; 4.5]	0.845	0.939	-4.0 [-7.8 ; -0.1]	0.046	0.116
miR-132-3p	-0.5 [-5.9 ; 5.2]	0.864	0.864	7.0 [1.5 ; 12.8]	0.013	0.064
miR-146a-5p	-2.9 [-8.6 ; 3.1]	0.337	0.842	-2.8 [-8.5 ; 3.3]	0.359	0.449
miR-222-3p	-2.6 [-7.4 ; 2.5]	0.310	0.999	-0.2 [-4.8 ; 4.7]	0.948	0.948
miR-223-3p	-1.4 [-5.5 ; 2.8]	0.505	0.842	-4.7 [-8.7 ; -0.4]	0.031	0.103
miR-451a	6.0 [-1.7 ; 14.2]	0.132	0.659	11.3 [3.7 ; 19.6]	0.003	0.033
miR-93-5p	2.2 [-1.4 ; 6]	0.227		5.2 [1.6 ; 8.9]	0.004	
miR-191-5p	-2.2 [-5.7 ; 1.4]	0.227		-4.9 [-8.2 ; -1.6]	0.004	

* False Discovery Rate corrected p value
Adjusted for age, sex, Body mass index (BMI) and year of inclusion

Supplemental table 3 : Association between residential NO₂ and miRNA using miRNA minus- average of the two housekeeping micro-ARN

micro-RNA	Men			Women				
	NO ₂ (per 5µg/m ³ increase)			NO ₂ (per 5µg/m ³ increase)				
	% of variation [IC95%]	p	p FDR	% of variation [IC95%]	p	p FDR*	p interaction	P interaction FDR
miR-21-5p	0.1 [-3.4 ; 3.7]	0,976	0,976	-1.4 [-5.4 ; 2.6]	0,483	0,536	0,577	0,962
miR-23a-5p	-3.0 [-7.7 ; 2.0]	0,236	0,589	-4.9 [-9.8 ; 0.3]	0,065	0,093	0,653	0,725
miR-28-3p	-5.6 [-9.6 ; -1.3]	0,011	0,105	-2.0 [-6.4 ; 2.6]	0,383	0,479	0,253	0,506
miR-30b-5p	0.2 [-4.3 ; 5.0]	0,926	0,999	-5.0 [-9.7 ; -0.2]	0,043	0,086	0,101	0,507
miR-126-3p	-0.3 [-3.6 ; 3.1]	0,867	0,999	-3.9 [-7.2 ; -0.5]	0,024	0,121	0,128	0,428
miR-132-3p	-1.4 [-5.9 ; 3.3]	0,551	0,787	4.9 [0.2 ; 9.8]	0,041	0,103	0,046	0,461
miR-146a-5p	-3.2 [-7.9 ; 1.9]	0,214	0,714	-5.3 [-10.1 ; -0.2]	0,041	0,135	0,636	0,795
miR-222-3p	-2.1 [-6.2 ; 2.1]	0,313	0,522	-1.1 [-5.1 ; 3.0]	0,581	0,581	0,630	0,900
miR-223-3p	-1.9 [-5.3 ; 1.6]	0,284	0,568	-5.3 [-8.8 ; -1.7]	0,005	0,048	0,158	0,394
miR-451a	4.4 [-2.0 ; 11.1]	0,183	0,915	6.6 [0.2 ; 13.4]	0,045	0,075	0,677	0,677
miR-93-5p	1.9 [-1.1 ; 5.0]	0,222		5.0 [1.9 ; 8.2]	0,001		0,189	
miR-191-5p	-1.9 [-4.8 ; 1.1]	0,222		-4.8 [-7.6 ; -1.9]	0,001		0,189	

* False Discovery Rate corrected p value
Adjusted for age, sex, Body mass index (BMI) and year of inclusion

Supplemental table 4 : Association between residential PM₁₀ and NO₂ and miRNA, using the level of the miRNA minus the average of the two housekeeping micro-ARN and after the exclusion of samples with hemolysis

PM ₁₀ (per 2 µg/m ³ increment)				NO ₂ (per 5 µg/m ³ increment)			
micro-RNA	% difference [95CI%]	p	p FDR	micro-RNA	% difference [95CI%]	p	p FDR*
miR.451a	8.8 [3.6 ; 14.3]	0,001	0,009	miR.28-3p	-3.8 [-6.6 ; -0.9]	0,010	0,104
miR.28-3p	-5.0 [-8.3 ; -1.7]	0,004	0,018	miR.223-3p	-3.1 [-5.4 ; -0.7]	0,013	0,065
miR.223-3p	-2.8 [-5.6 ; 0.0]	0,051	0,17	miR.146a-5p	-4.1 [-7.4 ; -0.8]	0,017	0,056
miR.146a-5p	-3.5 [-7.3 ; 0.6]	0,091	0,226	miR.451a	5.1 [0.8 ; 9.6]	0,021	0,052
miR.23a-5p	-3.3 [-7.0 ; 0.7]	0,104	0,208	miR.126-3p	-2.2 [-4.4 ; 0.1]	0,067	0,134
miR.126-3p	-2.1 [-4.8 ; 0.6]	0,124	0,206	miR.23a-5p	-3.1 [-6.3 ; 0.3]	0,070	0,117
miR.132-3p	2.7 [-1.3 ; 6.9]	0,185	0,264	miR.30b-5p	-2.1 [-5.4 ; 1.4]	0,245	0,350
miR.30b-5p	-1.7 [-5.7 ; 2.4]	0,405	0,506	miR.222-3p	-1.5 [-4.5 ; 1.6]	0,330	0,413
miR.222-3p	-1.5 [-5.0 ; 2.2]	0,421	0,468	miR.132-3p	1.7 [-1.7 ; 5.2]	0,332	0,369
miR.21-5p	0.2 [-3.0 ; 3.6]	0,889	0,889	miR.21-5p	-0.6 [-3.3 ; 2.3]	0,699	0,699
miR.93-5p	3.4 [1.1 ; 5.7]	0,004		miR.93-5p	2.8 [0.9 ; 4.8]	0,004	
miR.191-5p	-3.3 [-5.4 ; -1.1]	0,004		miR.191-5p	-2.8 [-4.6 ; -0.9]	0,004	

Supplemental table 5 : Association between residential PM₁₀ and NO₂ and miRNA, using the level of the miRNA minus the average of the three most stable micro-ARN.

PM ₁₀ (per 2 µg/m ³ increment)				NO ₂ (per 5 µg/m ³ increment)			
micro-RNA	% difference [95CI%]	p	p FDR	micro-RNA	% difference [95CI%]	p	p FDR*
miR.451a	9.7 [3.8 ; 15.9]	0,001	0.010	miR.451a	6.7 [1.8 ; 11.9]	0,007	0,063
miR.132-3p	4.2 [0.6 ; 7.9]	0,022	0,098	miR.146a-5p	-3.0 [-5.8 ; -0.2]	0,038	0,169
miR.28-3p	-3.3 [-6.1 ; -0.4]	0,029	0,086	miR.223-3p	-2.3 [-4.5 ; -0.1]	0.040	0.120
miR.223-3p	-2.1 [-4.6 ; 0.5]	0,109	0,245	miR.28-3p	-2.6 [-5 ; 0.0]	0,046	0,104
miR.146a-5p	-2.0 [-5.3 ; 1.4]	0,248	0,446	miR.132-3p	2.9 [-0.1 ; 6.0]	0,059	0,106
miR.21-5p	1.5 [-1.1 ; 4.2]	0,271	0,407	miR.126-3p	-0.9 [-2.8 ; 1.1]	0,397	0,595
miR.126-3p	-1.0 [-3.3 ; 1.4]	0,419	0,538	miR.30b-5p	-1.0 [-3.9 ; 2.0]	0,526	0,677
miR.30b-5p	-0.7 [-4.1 ; 2.9]	0,705	0,793	miR.21-5p	0.6 [-1.6 ; 2.8]	0,607	0,682
miR.222-3p	-0.6 [-3.6 ; 2.5]	0,709	0,709	miR.222-3p	-0.5 [-3.1 ; 2.1]	0,686	0,686
miR.23a-5p	-2.0 [-4.8 ; 0.8]	0,166		miR.23a-5p	-2.7 [-5 ; -0.3]	0,029	
miR.93-5p	4.8 [1.3 ; 8.5]	0,007		miR.93-5p	4.9 [1.9 ; 8.0]	0,001	
miR.191-5p	-2.7 [-4.7 ; -0.5]	0,014		miR.191-5p	-2.0 [-3.8 ; -0.2]	0,028	

* p value corrected for the false discovery rate

Adjusted for age, sex, body mass index, and year of inclusion

PM₁₀: particulate matter <10 µm, NO₂: nitrogen dioxide

Supplemental table 6: Association between residential PM₁₀ and NO₂ and miRNA, using the level of the miRNA minus the average of all the other micro-ARN.

PM ₁₀ (per 2 µg/m ³ increment)				NO ₂ (per 5 µg/m ³ increment)			
micro-RNA	% difference [95CI%]	p	p FDR	micro-RNA	% difference [95CI%]	p	p FDR*
miR.451a	10.2 [4.0 ; 16.8]	0,001	0,012	miR.451a	7.7 [2.5 ; 13.1]	0,003	0,041
miR.191-5p	-3.4 [-5.7 ; -0.9]	0,008	0,046	miR.93-5p	5.3 [1.7 ; 9.1]	0,004	0,024
miR.28-3p	-3.8 [-6.8 ; -0.8]	0,015	0,059	miR.146a-5p	-3.4 [-6.2 ; -0.6]	0,019	0,074
miR.93-5p	5.0 [0.8 ; 9.5]	0,021	0,062	miR.23a-5p	-3.0 [-5.6 ; -0.4]	0,027	0,081
miR.132-3p	4.0 [0.3 ; 7.8]	0,036	0,086	miR.223-3p	-2.3 [-4.5 ; -0.1]	0,040	0,097
miR.223-3p	-2.7 [-5.2 ; -0.1]	0,044	0,089	miR.28-3p	-2.7 [-5.3 ; -0.1]	0,041	0,081
miR.146a-5p	-3.0 [-6.2 ; 0.4]	0,079	0,136	miR.191-5p	-2.1 [-4.2 ; 0.0]	0,052	0,089
miR.23a-5p	-2.8 [-5.8 ; 0.3]	0,081	0,122	miR.132-3p	3.1 [0.0 ; 6.3]	0,052	0,078
miR.126-3p	-1.3 [-3.5 ; 0.9]	0,237	0,316	miR.126-3p	-0.8 [-2.7 ; 1.1]		0,534
miR.21-5p	1.1 [-1.2 ; 3.5]	0,350	0,420	miR.21-5p	0.8 [-1.2 ; 2.8]	0,448	0,537
miR.222-3p	-1.3 [-4.3 ; 1.7]	0,391	0,427	miR.222-3p	-0.9 [-3.5 ; 1.6]	0,471	0,514
miR.30b-5p	-1.1 [-4.4 ; 2.4]	0,539	0,539	miR.30b-5p	-0.8 [-3.7 ; 2.1]	0,570	0,570

* p value corrected for the false discovery rate
 Adjusted for age, sex, body mass index, and year of inclusion
 PM₁₀: particulate matter <10 µm, NO₂: nitrogen dioxide

Supplemental table 7 : description of previous study on association between air pollution and miR-21-5p, miR-23a-5p, miR-28-3p, miR-30b-5p, miR-126-3p, miR-132-3p, miR-146a-5p, miR-222-3p, miR-223-3p, miR-451a, miR-93-5p or miR-191-5p.

Author	year	Country	Population	Number of subjects	studied exposure	methodology	length of exposure	miRNA studied in common with our study
Tsamou (1)	2018	Belgium	pregnant women	210	PM _{2.5}	Cohort	exposure during pregnancy	miR-21,miR-146a,miR-222,in placenta at birth
Rider (2)	2016	Canada	Non-smoker	15	experimental exposure to diesel exhaust	Cross-over	short term	800 miRNA(bronchial brushings)
DU (3)	2022	Shanghai (China)	healthy college student	35	road session vs park session (TRAP)	Cross-over	short term	all
Chen (4)	2018	Shanghai (China)	student	55	true vs sham air purifier : PM _{2.5}	Cross-over	short term (9-days)	miR-21.5p,miR-146a-5p miR-93-5p,
Krauskopf (5)	2018	London (UK)	non-smoker. mean age 65	24	PM ₁₀ , PM _{2.5} ,black carbon, UFP, NO ₂	Cross-over	short term (2-hours)	miR-30.5p,miR-223.3p
Krauskopf (6)	2019	Barcelona (spain)	Healthy non-smokers aged from 18 to 60 years	24	PM ₁₀ , PM _{2.5} ,UFP, BC ,NO ₂ , CO, CO ₂	Cross-over	Short term (7h30)	miR-28-3p, miR-30b-5p, miR-222-3p, miR-146a-5p
Hou (7)	2016	Beijing (china)	60 truck driver and 60 office worker	120	PM _{2.5} , EC	repeated measurement	Short term	miR-146a
Vriens (8)	2016	Belgium	childrens	80	PM _{2.5} , UFP	repeated measurement	short term	miR-146a (saliva)
Mancini (9)	2020	Switzerland, United Kingdom, Italy , Netherlands	heathly non-smoking adults	143	PM _{2.5}	repeated measurment	short term 24-hours	none

Supplemental table 7 (continued) : description of previous study on association between air pollution and miR-21-5p, miR-23a-5p, miR-28-3p, miR-30b-5p, miR-126-3p, miR-132-3p, miR-146a-5p, miR-222-3p, miR-223-3p, miR-451a, miR-93-5p or miR-191-5p.

Author	year	Country	Population	Number of subjects	studied exposure	methodology	length of exposure	miRNA studied common with our study
Chen (10)	2020	North Carolina (USA)	coronary artery disease patients	14	Ozone, PM _{2.5}	repeated measurement	short term (1 to 5-days)	miR-21-5p, miR-126-5p, miR-146-5p,
Louwies (11)	2016	Belgium	helthy adults	50	PM ₁₀	repeated measurement	short term (2-hours to 1-week)	miR-21, miR-146a, miR-222
Rodosthenous (12)	2016	Boston (USA)	men between 21 and 80 years	22	PM _{2.5}	repeated measurement	short term (1 day) intermediate (1-week, 1-month) long term (6-month, 1-year)	miR23a-3p, miR93-5p, miR-126-3p, miR-146a-5p, miR-191-5p, miR-223-3p, miR-451a
Fossati (13)	2013	Boston (USA)	Elderly men "Normative Aging Study"	153	PM _{2.5} , BC	Cross sectional	short term - intermediate (4-hours to 28-days)	miR.21, miR.126, miR.146a, miR.222
Motta (14)	2016	Italy	Obese and overweight	90	PM ₁₀	Cross sectional	short term	miR-21, miR-30b, miR-222, miR-451
Li (15)	2020	Jinan China	Childrens	273	(BPDE-Alb) biomarker of PM exposure	Cross sectional	long term	miR-21-5p, miR-146a-5p

BPDE-Alb : benzo[a]pyrene-r-7,t-8,t-9,c-10-tetrahydrotetrol and serum albumin ; PM : particulate matter ; PM_{2.5} particulate matter <2.5 µm; PM₁₀: particulate matter <10 µm ; NO₂: nitrogen dioxide; BC : Black Carbon, EC : elemental carbon ; UFC : Ultrafine particle; CO carbon monoxide; CO₂ Carbon dioxide ; TRAP: traffic air pollution.

Supplemental table 8: Significant association observed in literature between air pollution and miR-21-5p, miR-23a-5p, miR-28-3p, miR-30b-5p, miR-126-3p, miR-132-3p, miR-146a-5p, miR-222-3p, miR-223-3p, miR-451a, miR-93-5p or miR-191-5p

MiRNA Main Biological effect	Biological effect	Long term exposure		Short term exposure	
		Exposure Positively associated with miRNA	Exposure negatively associated with miRNA	Exposure Positively associated with miRNA	Exposure negatively associated with miRNA
miR-21-5p Inflammation	Inhibition of tumor suppressor genes such as PTEN and PDCD4 (16) Positive regulation of the NF-κB/NLRP3 pathways (17)	BPDE-Alb ; li (15)	NO ₂ and PM _{2.5} exposure during trimester 2 of pregnancy ; Tsamou (1)	Controlled diesel exhaust ; Rider (2)	PM _{2.5} ; Chen 2018 (4) PM _{2.5} , BC ; Fossati (2) PM ₁₀ ; Louwies (11) TRAP ; DU (3)
miR-23a-5p inflammation	Inhibition of NLRP3-activating factors, (18,19)	PM10 (6 month 1 year) Rodosthenous (12)	NO ₂ ; Hubert		TRAP ; DU (3)
miR-28-3p inflammation	Downregulation of the NRF2 signaling pathway, (20)		NO ₂ ; Hubert	TRAP; Krauskopf 2019. (6)	
miR-30b-5p inflammation	Regulation of the JAK/STAT3 pathway through inhibition of SOCS3, (21)			TRAP ; Krauskopf 2019 (6)	PM ₁₀ ; Motta(14) TRAP ; DU (3)
miR-126-3p carcinogenesis	Restriction of cell proliferation via inhibiting PLXNB2 expression and the subsequent RhoA/ROCK signaling pathway.(22) Regulation of the PIK3/AKT signaling cascade by inhibition of IRS-1, PIK3R2 and AKT2. (23–25)	PM2.5 (6 month ,1 year) ; Rodosthenous (12)		PM ₁₀ ; Rodosthenous(12)	PM _{2.5} ; BC ; Fossati (13)
miR-132-3p inflammation	Upregulation of inflammatory and apoptosis by targeting SIRT1, (25)				TRAP ; Krauskopf 2018(5)

Supplemental table 8 (continued) : Significant association observed in literature between air pollution and miR-21-5p, miR-23a-5p, miR-28-3p, miR-30b-5p, miR-126-3p, miR-132-3p, miR-146a-5p, miR-222-3p, miR-223-3p, miR-451a, miR-93-5p or miR-191-5p

MiRNA Main Biological effect	Biological effect	Long term exposure		Short term exposure	
		Exposure Positively associated with miRNA	Exposure negatively associated with miRNA	Exposure Positively associated with miRNA	Exposure negatively associated with miRNA
miR-146a-5p inflammation	Downregulation of the NF- κ B-mediated priming of the NLRP3 inflammasome, (26,27)	BPDE-Alb; li (15)	NO ₂ , Hubert	TRAP ; Krauskopf 2019 (6) PM ₁₀ ; Rodosthenous(12)	PM _{2.5} ; Chen 2018 (4) PM _{2.5} , BC ; Fossati (13) NO ₂ ; Chen 2020 (10)
miR-222-3p inflammation	Modulation of the JAK/STAT pathways by targeting SOCS1 (29,30)		PM _{2.5} exposure during trimester 2 of pregnancy (1)	TRAP ; Krauskopf 2019 (6) UFP ; Vriens (8)	PM _{2.5} , BC ; Fossati (2) PM ₁₀ ; Louwies (11) TRAP ; DU (3)
miR-223-3p inflammation	Inhibition of the NLRP3 inflammasome (31,32)	PM _{2.5} (6 month and 1 year) ; Rodosthenous (12)	NO ₂ , Hubert	PM ₁₀ , Rodosthenous (12) Controlled diesel exhaust ; Rider (2)	
miR-451a inflammation	Inhibition of the NF-kB pathway (33–36)	PM ₁₀ ,NO ₂ ; Hubert			Rodosthenous (12)
miR-93-5p (endogenous control) inflammation	Regulation of IL-8 and VEGF gene expression (37) Involvement in oxidative stress-induced mitophagy (38)	PM ₁₀ , NO ₂ ;Hubert PM _{2.5} (6 month and 1 year) Rodosthenous(12)			
miR-191-5p (endogenous control) inflammation	Downregulation of the NLRP3 inflammasome by repressing C/EBP β , (39); Inhibition of the MAPK signaling pathway by targeting Map3k12, (40)	PM _{2.5} (6 month and 1 year) Rodosthenous (12)	PM ₁₀ , NO ₂ ; Hubert		

1. Tsamou M, Vrijens K, Madhloum N, Lefebvre W, Vanpoucke C, Nawrot TS. Air pollution-induced placental epigenetic alterations in early life: a candidate miRNA approach. *Epigenetics*. 2018;13(2):135-46.
2. Rider CF, Yamamoto M, Günther OP, Hirota JA, Singh A, Tebbutt SJ, et al. Controlled diesel exhaust and allergen coexposure modulates microRNA and gene expression in humans: Effects on inflammatory lung markers. *Journal of Allergy and Clinical Immunology*. déc 2016;138(6):1690-700.
3. Du X, Zhang Q, Jiang Y, Zhu X, Zhang Y, Liu C, et al. Characterization of plasma-derived exosomal miRNA changes following traffic-related air pollution exposure: A randomized, crossover trial based on small RNA sequencing. *Environment International*. sept 2022;167:107430.
4. Chen R, Li H, Cai J, Wang C, Lin Z, Liu C, et al. Fine Particulate Air Pollution and the Expression of microRNAs and Circulating Cytokines Relevant to Inflammation, Coagulation, and Vasoconstriction. *Environ Health Perspect*. 17 janv 2018;126(1):017007.
5. Krauskopf J, Caiment F, van Veldhoven K, Chadeau-Hyam M, Sinharay R, Chung KF, et al. The human circulating miRNome reflects multiple organ disease risks in association with short-term exposure to traffic-related air pollution. *Environ Int*. avr 2018;113:26-34.
6. Krauskopf J, van Veldhoven K, Chadeau-Hyam M, Vermeulen R, Carrasco-Turigas G, Nieuwenhuijsen M, et al. Short-term exposure to traffic-related air pollution reveals a compound-specific circulating miRNA profile indicating multiple disease risks. *Environ Int*. juill 2019;128:193-200.
7. Hou L, Barupal J, Zhang W, Zheng Y, Liu L, Zhang X, et al. Particulate Air Pollution Exposure and Expression of Viral and Human MicroRNAs in Blood: The Beijing Truck Driver Air Pollution Study. *Environ Health Perspect*. mars 2016;124(3):344-50.
8. Vriens A, Nawrot TS, Saenen ND, Provost EB, Kicinski M, Lefebvre W, et al. Recent exposure to ultrafine particles in school children alters miR-222 expression in the extracellular fraction of saliva. *Environ Health*. 26 juill 2016;15(1):80.
9. Mancini FR, Laine JE, Tarallo S, Vlaanderen J, Vermeulen R, van Nunen E, et al. microRNA expression profiles and personal monitoring of exposure to particulate matter. *Environ Pollut*. août 2020;263(Pt B):114392.
10. Chen H, Xu Y, Rappold A, Diaz-Sanchez D, Tong H. Effects of ambient ozone exposure on circulating extracellular vehicle microRNA levels in coronary artery disease patients. *Journal of Toxicology and Environmental Health, Part A*. 2 mai 2020;83(9):351-62.
11. Louwies T, Vuegen C, Panis LI, Cox B, Vrijens K, Nawrot TS, et al. miRNA expression profiles and retinal blood vessel calibers are associated with short-term particulate matter air pollution exposure. *Environ Res*. mai 2016;147:24-31.
12. Rodosthenous RS, Coull BA, Lu Q, Vokonas PS, Schwartz JD, Baccarelli AA. Ambient particulate matter and microRNAs in extracellular vesicles: a pilot study of older individuals. *Part Fibre Toxicol*. 8 mars 2016;13:13.
13. Fossati S, Baccarelli A, Zanobetti A, Hoxha M, Vokonas PS, Wright RO, et al. Ambient particulate air pollution and microRNAs in elderly men. *Epidemiology*. janv 2014;25(1):68-78.
14. Motta V, Favero C, Dioni L, Iodice S, Battaglia C, Angelici L, et al. MicroRNAs are associated with blood-pressure effects of exposure to particulate matter: Results from a mediated moderation analysis. *Environ Res*. avr 2016;146:274-81.
15. Li J, Wang T, Wang Y, Xu M, Zhang L, Li X, et al. Particulate matter air pollution and the expression of microRNAs and pro-inflammatory genes: Association and mediation among children in Jinan, China. *J Hazard Mater*. 5 mai 2020;389:121843.
16. Matsushashi S, Manirujjaman M, Hamajima H, Ozaki I. Control Mechanisms of the Tumor Suppressor PDCD4: Expression and Functions. *IJMS*. 9 mai 2019;20(9):2304.

17. Xue Z, Xi Q, Liu H, Guo X, Zhang J, Zhang Z, et al. miR-21 promotes NLRP3 inflammasome activation to mediate pyroptosis and endotoxic shock. *Cell Death Dis.* 12 juin 2019;10(6):461.
18. Chang H, Chang H, Cheng T, Lee GD, Chen X, Qi K. Micro-ribonucleic acid-23a-3p prevents the onset of type 2 diabetes mellitus by suppressing the activation of nucleotide-binding oligomerization-like receptor family pyrin domain containing 3 inflammatory bodies-caused pyroptosis through negatively regulating NIMA-related kinase 7. *J Diabetes Investig.* mars 2021;12(3):334-45.
19. Pan Z, Shan Q, Gu P, Wang XM, Tai LW, Sun M, et al. miRNA-23a/CXCR4 regulates neuropathic pain via directly targeting TXNIP/NLRP3 inflammasome axis. *J Neuroinflammation.* 31 janv 2018;15(1):29.
20. Yang M, Yao Y, Eades G, Zhang Y, Zhou Q. MiR-28 regulates Nrf2 expression through a Keap1-independent mechanism. *Breast Cancer Res Treat.* oct 2011;129(3):983-91.
21. Zhou T, Chen Y li. The Functional Mechanisms of miR-30b-5p in Acute Lung Injury in Children. *Med Sci Monit.* 2 janv 2019;25:40-51.
22. Chen Q, Hu H, Jiao D, Yan J, Xu W, Tang X, et al. miR-126-3p and miR-451a correlate with clinicopathological features of lung adenocarcinoma: The underlying molecular mechanisms. *Oncology Reports.* août 2016;36(2):909-17.
23. Guo C, Sah JF, Beard L, Willson JKV, Markowitz SD, Guda K. The noncoding RNA, miR-126, suppresses the growth of neoplastic cells by targeting phosphatidylinositol 3-kinase signaling and is frequently lost in colon cancers. *Genes Chromosom Cancer.* nov 2008;47(11):939-46.
24. Zhang J, Du Y ying, Lin Y feng, Chen Y ting, Yang L, Wang H jun, et al. The cell growth suppressor, mir-126, targets IRS-1. *Biochemical and Biophysical Research Communications.* déc 2008;377(1):136-40.
25. Sibilano M, Tullio V, Adorno G, Savini I, Gasperi V, Catani MV. Platelet-Derived miR-126-3p Directly Targets AKT2 and Exerts Anti-Tumor Effects in Breast Cancer Cells: Further Insights in Platelet-Cancer Interplay. *IJMS.* 13 mai 2022;23(10):5484.
26. Han S, Lin F, Ruan Y, Zhao S, Yuan R, Ning J, et al. miR-132-3p promotes the cisplatin-induced apoptosis and inflammatory response of renal tubular epithelial cells by targeting SIRT1 via the NF- κ B pathway. *International Immunopharmacology.* oct 2021;99:108022.
27. Olivieri F, Prattichizzo F, Giuliani A, Matakchione G, Rippo MR, Sabbatinelli J, et al. miR-21 and miR-146a: The microRNAs of inflammaging and age-related diseases. *Ageing Res Rev.* sept 2021;70:101374.
28. Hou J, Deng Q, Deng X, Zhong W, Liu S, Zhong Z. MicroRNA-146a-5p alleviates lipopolysaccharide-induced NLRP3 inflammasome injury and pro-inflammatory cytokine production via the regulation of TRAF6 and IRAK1 in human umbilical vein endothelial cells (HUVECs). *Ann Transl Med.* sept 2021;9(18):1433.
29. Department of Gastroenterology, The Affiliated Jiangning Hospital of Nanjing Medical University, Jiangsu, China, Xia F, Bo W, Department of Gastroenterology, The Affiliated Jiangning Hospital of Nanjing Medical University, Jiangsu, China, Ding J, Department of Gastroenterology, The Affiliated Jiangning Hospital of Nanjing Medical University, Jiangsu, China, et al. MiR-222-3p Aggravates the Inflammatory Response by Targeting SOCS1 to Activate STAT3 Signaling in Ulcerative Colitis. *Turk J Gastroenterol.* 21 nov 2022;33(11):934-44.
30. Zhang P, Yu J, Gui Y, Sun C, Han W. Inhibition of miRNA-222-3p Relieves Staphylococcal Enterotoxin B-Induced Liver Inflammatory Injury by Upregulating Suppressors of Cytokine Signaling 1. *Yonsei Med J.* 2019;60(11):1093.
31. Bauernfeind F, Rieger A, Schildberg FA, Knolle PA, Schmid-Burgk JL, Hornung V. NLRP3 inflammasome activity is negatively controlled by miR-223. *J Immunol.* 15 oct 2012;189(8):4175-81.

32. Haneklaus M, Gerlic M, Kurowska-Stolarska M, Rainey AA, Pich D, McInnes IB, et al. Cutting edge: miR-223 and EBV miR-BART15 regulate the NLRP3 inflammasome and IL-1 β production. *J Immunol.* 15 oct 2012;189(8):3795-9.
33. Hur W, Lee JH, Kim SW, Kim JH, Bae SH, Kim M, et al. Downregulation of microRNA-451 in non-alcoholic steatohepatitis inhibits fatty acid-induced proinflammatory cytokine production through the AMPK/AKT pathway. *Int J Biochem Cell Biol.* juill 2015;64:265-76.
34. Li HP, Zeng XC, Zhang B, Long JT, Zhou B, Tan GS, et al. miR-451 inhibits cell proliferation in human hepatocellular carcinoma through direct suppression of IKK- β . *Carcinogenesis.* nov 2013;34(11):2443-51.
35. Xie J, Hu X, Yi C, Hu G, Zhou X, Jiang H. MicroRNA-451 protects against cardiomyocyte anoxia/reoxygenation injury by inhibiting high mobility group box 1 expression. *Mol Med Rep.* juin 2016;13(6):5335-41.
36. Barnay-Verdier S, Maréchal V, Borde C. [HMGB1: a link between innate and adaptive immunity]. *Rev Francoph Lab.* déc 2009;2009(417):59-68.
37. Fabbri E, Montagner G, Bianchi N, Finotti A, Borgatti M, Lampronti I, et al. MicroRNA miR-93-5p regulates expression of IL-8 and VEGF in neuroblastoma SK-N-AS cells. *Oncology Reports.* mai 2016;35(5):2866-72.
38. Zhang C, Nie P, Zhou C, Hu Y, Duan S, Gu M, et al. Oxidative stress-induced mitophagy is suppressed by the miR-106b-93-25 cluster in a protective manner. *Cell Death Dis.* 24 févr 2021;12(2):209.
39. Chen J, Sun J, Hu Y, Wan X, Wang Y, Gao M, et al. MicroRNA-191-5p ameliorates amyloid- β ₁₋₄₀-mediated retinal pigment epithelium cell injury by suppressing the NLRP3 inflammasome pathway. *The FASEB Journal [Internet].* avr 2021 [cité 2 mars 2023];35(4). Disponible sur: <https://onlinelibrary.wiley.com/doi/10.1096/fj.202000645RR>
40. Wan W, Liu G, Li X, Liu Y, Wang Y, Pan H, et al. MiR-191-5p alleviates microglial cell injury by targeting Map3k12 (mitogen-activated protein kinase kinase kinase 12) to inhibit the MAPK (mitogen-activated protein kinase) signaling pathway in Alzheimer's disease. *Bioengineered.* 20 déc 2021;12(2):12678-90.

AUTEURE : Nom : HUBERT

Prénom : Audrey

Date de soutenance : 20/09/2024

Titre de la thèse : La relation entre l'exposition résidentielle à la pollution atmosphérique et les micro-ARN circulants chez les adultes vivant en zone urbaine du nord de la France

Thèse - Médecine - Lille 2024

Cadre de classement : *Médecine*

DES: DES Santé Publique et Médecine Sociale

Mots-clés : Santé environnementale, pollution atmosphérique, particules en suspension, dioxyde d'azote, micro-ARN, biomarqueurs

Résumé :

Introduction : Les micro-ARN sont des facteurs de régulation épigénétique capables de réprimer l'expression des gènes cibles et pourraient jouer un rôle dans les effets de la pollution atmosphérique sur la santé. L'objectif de la présente étude en population générale était d'examiner l'association entre l'expression des micro-ARN et l'exposition résidentielle à long terme aux PM₁₀ et au NO₂.

Méthode : Nous avons inclus 998 participants adultes non-fumeurs issus de l'enquête transversale ELISABET (2010-2014) dans la région urbaine de Lille, en France. Les niveaux moyens annuels de pollution résidentielle ont été estimés à l'aide d'un système de modélisation de dispersion atmosphérique. Dix micro-ARN ont été sélectionnés sur la base des données disponibles dans la littérature, ainsi que deux micro-ARN de référence (miR-93-5p et miR-191-5p), et ont été quantifiés par RT-qPCR. Des modèles de régression linéaire multivariée ont été utilisés pour étudier l'association entre les micro-ARN et la pollution atmosphérique. Le seuil de signification statistique (après correction pour le taux de fausse découverte) a été fixé à $p < 0,1$.

Résultats : L'exposition annuelle moyenne entre 2011 et l'année d'inclusion était de $26,4 \pm 2,0$ $\mu\text{g}/\text{m}^3$ pour les PM₁₀ et de $24,7 \pm 5,1$ $\mu\text{g}/\text{m}^3$ pour le NO₂. Chaque augmentation de 2 $\mu\text{g}/\text{m}^3$ d'exposition aux PM₁₀ était associée à une augmentation de 8,6 % (IC 95 % [3,1 ; 14,3]; $p_{\text{FDR}} = 0,019$) de l'expression de miR-451a. Une augmentation de 5 $\mu\text{g}/\text{m}^3$ de l'exposition au NO₂ était associée à une augmentation de 5,3 % (IC 95 % [0,7 ; 10]; $p_{\text{FDR}} = 0,056$) de l'expression de miR-451a, une diminution de 3,6 % (IC 95 % [-6,1 ; -1,1]; $p_{\text{FDR}} = 0,052$) de l'expression de miR-223-3p, une diminution de 3,8 % (IC 95 % [-6,8 ; -0,7]; $p_{\text{FDR}} = 0,079$) de l'expression de miR-28-3p, une diminution de 4,3 % (IC 95 % [-7,7 ; -0,8]; $p_{\text{FDR}} = 0,055$) de l'expression de miR-146a-5p et une diminution de 4,0 % (IC 95 % [-7,4 ; -0,4]; $p_{\text{FDR}} = 0,059$) de l'expression de miR-23a-5p. La différence entre les deux micro-ARN de référence miR-93-5p et miR-191-5p était également associée à l'exposition aux PM₁₀ et au NO₂.

Conclusion : Nos résultats suggèrent que les micro-ARN circulants pourraient être des biomarqueurs précieux pour évaluer les effets de la pollution atmosphérique.

Composition du Jury :

Président : Monsieur le Professeur Philippe AMOUYEL

Assesseurs : Monsieur le Docteur Jean-Marc LO GUIDICE

Madame la Docteur Victoria GAUTHIER

Directeur de thèse : Monsieur le Docteur Luc DAUCHET